

Review Article

Chemical Constituents of Loranthaceae (Mistletoes): A Review

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ABSTRACT

This article review is designed to bring some updates on the phytochemistry of the Loranthaceae family in order to throw more light on future research priorities. Loranthaceae family is the leading pantropical semi parasitic shrubs generally termed mistletoe. They consist of about 75 genera and 1000 species growing in many countries of Africa, Asia, Australia, Europe, South America and New Zealand with diverse widespread uses in adornment, food and medication. The literatures for this review was collected from various online databases: Google Scholar, PubMed, Research gate, Science Direct, Scopus and web of Science for nine genera namely, *Globimetula*, *Scurrula*, *Dendrophoe*, *Loranthus*, *Phragmanthera*, *Tapinanthus*, *Taxillus*, *Tripodanthus* and *Viscum* based on the type of phytochemicals, popularity and environmental setting. Chemical studies of the Loranthaceae species have produced a number of important phytochemicals belonging to different classes. The recent investigations led to the discovery of several secondary metabolites such as alkaloids, flavonoids, phenolic compounds, lignan and neolignans and triterpenes. Hence this review represents a comprehensive assessment of the phytochemical studies carried out on Loranthaceae plants and could be a source of therapeutically beneficial products.

Keywords: Loranthaceae, phytochemicals, alkaloids, triterpenes, flavonoids

1. INTRODUCTION

Medicinal plants are composed of certain substances that are active in curing and preventing precise ailments and diseases. They have played a fundamental role as the principal source of medicine in diverse ancient traditional system of medication either additively, separately, or in synergy for the treatment of various ailments (Elujoba, 1997). According to the World Health Organization (WHO), wrong handling of orthodox medicines or practices can have undesirable or dangerous effects and that further investigation is necessary to establish the effectiveness and safety of several medicinal plants used by traditional medicine system (WHO, 2014). The use of conventional medicine is conversely not limited to developing countries, surveys revealed that over 70% of population in developed countries like Canada and Germany have tried complementary or alternative medicine (CAM) at least once (Gurib-Fakim, 2006). Additionally, the growing reliance on the utilization of medicinal plants in advanced countries is substantiated by the extraction and evolution of many drugs and chemotherapeutics from these naturally occurring plants and conventionally used rural

medications (UNESCO, 1998). Medicinal plants have established their contributions to the management of ailments such as anaemia, cancer, malaria, HIV/AIDS, microbial infections, mental disorders, sickle-cell, and diabetes (Okigbo and Mbajiuka, 2005). They are composed of biologically active phytochemicals such as alkaloids, essential oils, flavonoids, saponins, tannins, triterpenes and other chemical compounds, which possess preventive and medicinal properties. These complex chemical constituents of diverse structural entities are found as secondary metabolites in one or more of these medicinal plants and are valuable to humankind. Explorations into the biological and pharmacological activities of medicinal plants throughout the past two centuries have produced compounds for the development of modern synthetic organic chemistry and the rise of medicinal chemistry as a main route for the discovery of new and more active natural products (Ogbulie et al., 2004). Natural products (secondary metabolites) are chemical compounds or substances found naturally from plants, aquatics and microbes that essentially have a biological or pharmacological activity for use in discovery and development of new drug. They are smaller molecular weight molecules below 3,000 Daltons produced by living organisms, plants, insects and have been the sources of most active ingredients of medications in the form of concoctions, lubricants, orthodox medicines and therapies with numerous bioactive compounds still being unidentified (Newman and Cragg, 2012; Kinghorn et al., 2009). The earliest known natural products records were depicted dated from 2600 BC on a Sumerian clay slabs which recorded oils from two species of plants that are still in use today as remedies for various ailments like colds, coughs and inflammations [9]. The Ebers papyrus is an aged and best known Egyptian Pharmaceutical records dating from about 2900 BC which listed hundreds of plants based medications (Cragg GM and Newman, 2009). The Chinese "Materia Medica" dated about 1100 BC well documented the use of thousands of plants derived natural products credited to China's famous ruler Shennong (Cragg GM and Newman, 2009). The Ayurvedic and Unanic medicinal systems of India recorded in Susruta and Charaka date from 1000 BC (Shoeb, 2006). The early Greek use of natural product can be traced back to Greek Legendary Doctor Dioscorides (100 A.D.) that described and documented the uses of more than 600 medicinal plants. His work remains the standard medical reference in most European countries for the over 1500 years (Cragg GM and Newman, 2009).

Therapeutic agents have been the most essential source of active compounds and chemical lead structures that led to the discovery and development of new drugs. Nevertheless, while many of the drugs used in the last 50 years or more have been of artificial origin, the pharmacopoeias earlier to that period were of natural origin (Jeffreys, 2005). Among the earliest important examples of medical treatment from natural sources include the discovery of salicin obtained in 1825-26 from the bark of white willow, *Salix Alba*. It was then improved to salicylic acid via hydrolysis and oxidation, and lastly to acetylsalicylic acid (ASA) via acetylation under the trade name aspirin in 1899 due to severe gastro intestinal toxicity of salicylic acid. Today, aspirin is still the most commonly used analgesic and antipyretic drug in the world (Jeffreys, 2005). Plant derived natural products such as Morphine and codeine isolated from the *Opium poppy* plant in 1804 have remained a rich source of lead compounds for effective pain killers (Beutler, 2009). The accidental discovery of antimalarial agent quinine from *Cinchona* led to the synthesis of many classes of antimalarial drugs based on quinine structure (Beutler, 2009). Clinically essential drugs isolated recently from plants include the anti-cancer agent paclitaxel (Taxol) and antimalarial agent artemisinin isolated from the yew tree and *Artemisia annua* respectively (Goodman, 2001).

Development of drugs from natural product sources have evolved over a lengthy period, and more than 50% of the most marketable drugs in clinical use were natural products or derivatives of natural products. It has been hypothesized that approximately 80% of drug constituents were natural products or enthused by a naturally occurring compound especially when drug discovery is considered before the introduction of high-throughput screening and

post genomic era. Natural products are originated from the occurrence of natural biodiversity in which the acquaintances between organisms and their ecosystem devise some diverse complex chemical entities inside the organisms that augment their subsistence and attractiveness (Harvey, 2008).

2. DISTRIBUTION, BOTANICAL DESCRIPTION AND ETHNOBOTANY

The family Loranthaceae is timbered flowering plant called mistletoes. They are hemiparasitic herbs or shrubs that attaches to a tree branches or aerial parts of the host plants by haustorium, an appendage through which they siphon water and minerals from the host plant, but photosynthesize their carbohydrates by means of their ever-green leaves. Loranthaceae is an essential plant with 75 genera and about 1000 species distributed throughout tropical regions (Burkhill, 1995). They are typically in widespread growing in Africa, Asia, America, Australia, Europe, South and New Zealand. Insistent stringy leaves and luminously coloured inflorescence largely characterized them. The leaves of the plants are opposite, displaced-opposite, rarely alternate, hairless, and sometimes petiolate or sessile. The flowers are bisexual and borne singly or in pairs. Their corollas are mostly red and splitting separately, the apical enlargement of the flower bud signifying maturity by changing to a darker colour; lobes twisted outwards at anthesis. The anthers are fitting below the top-shaped to peltate stigma. Their fruits are yellow or red berries with a luminously coloured seeds (Huaxing and Micheal, 2004). Members of this family parasitize a broad range of Gymnosperms and Angiosperms. Mistletoes are known all over the world to cause economic damages when infesting cultivated crops. They pose danger to plantation by parasitizing cultivated plants and tended plants (Burkhill, 1985).

Loranthaceae are agents of diseases and disruption and thus affect the host physiology leading to reduced growth, existence and reproduction (Gill and Onyibe, 1990; Kolb, 2002). Despite their destructive nature to their host, the plants are used medicinally for the treatment of many diseases including cardiovascular diseases (Ouedraogo et al., 2004), hepatic illness (Al-Ghaithi et al., 2004; Phillipson and Wright, 1991), fever and removal of placenta after parturition (Sher and Alyemeni, 2011), arthritis, hypertension and infertility (Matthes et al., 2005), abdominal pains, diabetes, fever and urinary tract infections (Adesina et al., 2013), anticancer, antihypertensive, antimicrobial and antioxidant (Ja'afar et al., 2017; Lim et al., 2016; Puneetha and Amruthesh, 2016), infusion for fatigue (Puneetha and Amruthesh, 2016), mental condition, fatigue, sterility, urino-genital problems, skin diseases, rickets, fractured limbs, rheumatism, coughs, malaria, chest conditions, infertility, stomach problems, and as a laxative (Ibrahim and Ayodele, 2013; Irvine, 1961; Musa et al., 2014; Olagunju et al., 1999; Traore et al., 2004), cholera, hypertension, diabetes, blood purifier, gastrointestinal tract diseases and wound infections (Deeni, 1989; Hussain and Karatela, 1989; Obatomi, 1994; Obatomi et al., 1996; Oliver, 1987). Scientific evidences have revealed that their compositions and medicinal activities are dependent on the host plant, harvesting period and species (Fukunaga et al., 1989; Oliver, 1987; Quan-Yu et al., 2015; Quan-Yu et al., 2016). Plants of similar family characteristically synthesize the same class of compounds due to the existence of similar enzymes and thus the same synthetic pathways (Bick, 1996).

3. CHEMICAL CONSTITUENTS

The phytochemical studies of various species of Loranthaceae have successfully afforded numerous chemical components. The customary uses of the plants in traditional medicine enthused the author to write on the chemical constituents of Loranthaceae family that led to the discovery of several secondary metabolites such as alkaloids, flavonoids, phenolic compounds,

lignans, neolignans and triterpenoids. As such, the chemistry of 9 genera *Globimetula*, *Scurrula*, *Dendrophoe*, *Loranthus*, *Phragmanthera*, *Tapinanthus*, *Taxillus*, *Tripodanthus* and *Viscum* have been studied and reported. The selection of these genera was based on nature of the compounds, popularity and geographical location. The phytochemical studies of Loranthaceae species together with their species and host plants are shown in Table 1.

Table 1. Summary of phytochemicals isolated from Loranthaceae species

Compounds	Species	Host Plant	References
ALKALOIDS			
Theobromine (1)	<i>S. atropurpurea</i>	<i>T. sinensis</i>	(Kazuyoshi et al., 2003)
Caffeine (2)	<i>S. atropurpurea</i>	<i>T. sinensis</i>	(Kazuyoshi et al., 2003)
4,5,4'-trihydroxy-3,3'-iminodibenzonic acid (3)	<i>V. album</i>	<i>P. nigra</i> , <i>P. x cana-densitis</i> <i>P. deltoides</i>	(Bashar et al., 2012)
4,5,4',5'-tetrahydroxy-3,3'-iminodibenzonic acid (4)	<i>V. album</i>	<i>P. nigra</i> , <i>P. x cana-densitis</i> <i>P. deltoides</i>	(Bashar et al., 2012)
Lupinine (5)	<i>L. micranthus</i>	<i>K. acuminata</i>	(Edwin et al., 2011)
FLAVONOIDS			
Quercetin 3-O- α -L-rhamnopyranoside (Quercitrin) (6)	<i>S. parasitica</i> <i>S. atropurpurea</i> , <i>D. falcate</i> <i>D. falcate</i> <i>S. ferruginea</i> <i>P. capitata</i> <i>L. kaoi</i> , <i>L. tanakae</i>	<i>N. indica</i> , <i>T. sinensis</i> , <i>S. robusta</i> <i>H. fomes</i> <i>NR</i> <i>C. spectabilis</i> <i>NR</i> <i>Quercus</i> and <i>Betula</i>	(Quan-Yu et al., 2015) (Kazuyoshi et al., 2003) (Mallavadhani et al., 2006) (Hasan et al., 2006) (Francoise et al., 2002) (Bruno et al., 2015) (Lin and Lin, 1999) (Young-Kyoong et al., 2004)
Naringin (7)	<i>V. album</i>	NR	(Yang et al., 2011)
Apigenin (8)	<i>G. braunii</i>	<i>P. thonningii</i>	(Ja'afar et al., 2017)
Apigenin-8-O- β -D(2"-O- β -D-glucopyranosyl)-glucopyranoside (9)	<i>G. braunii</i>	<i>P. thonningii</i>	(Muhammad et al., 2022)
Apigenin-8-C- β -d-glucopyranoside (10)	<i>P. capitata</i>	<i>P. insignis</i>	(Muhammad et al., 2019)
Kaempferol-3-O- α -L-rhamnopyranoside (11)	<i>D. falcate</i>	<i>S. robusta</i>	(Mallavadhani et al., 2006)
4"-O-acetylquercitrin (12)	<i>S. ferruginea</i>	NR	(Francoise et al., 2002)
Quercetin (13)	<i>S. ferruginea</i>	NR	(Francoise et al., 2002)
2-(2,4-dihydroxyphenyl)-5,7-dihydroxy-3-(3-methylhexyl)-4H-chromen-4-one (14)	<i>P. capitata</i> <i>T. theifer</i> <i>G. braunii</i> <i>G. braunii</i> <i>S. parasitica</i> <i>S. parasitica</i> <i>G. braunii</i> <i>P. austroarabica</i> <i>T. globiferus</i> <i>T. globiferus</i>	<i>P. spectabilis</i> <i>Z. serrata</i> <i>P. thonningii</i> <i>P. thonningii</i> <i>P. pinnata</i> <i>P. pinnata</i> <i>T. catappa</i> <i>NR</i> <i>V. doniana</i> <i>V. doniana</i>	(Bruno et al., 2015) (Tung-Hu et al., 2010) (Ja'afar et al., 2017) (Muhammad et al., 2022) (Muhammad et al., 2019) (Muhammad et al., 2019) (Danladi et al., 2010) (Goda et al., 2022) (Abubakar et al., 2020) (Abubakar et al., 2020)
Dihydromyricetin (15)	<i>S. liquidambaricola</i>	NR	(Shen et al., 1993)
Rutin (16)	<i>S. atropurpurea</i> , <i>T. acutifolius</i> <i>L. micranthus</i> <i>T. theifer</i> <i>G. braunii</i> <i>G. braunii</i> <i>S. parasitica</i> <i>S. atropurpurea</i> , <i>L. kaoi</i> <i>L. parasiticus</i>	<i>T. sinensis</i> , NR <i>H. brasiliensis</i> <i>Z. serrata</i> <i>P. thonningii</i> <i>P. thonningii</i> <i>N. indica</i> , <i>T. sinensis</i> , NR NR	(Kazuyoshi et al., 2003) (Soberón et al., 2010) (Matthias et al., 2014) (Tung-Hu et al., 2010) (Ja'afar et al., 2017) (Muhammad et al., 2022) (Quan-Yu et al., 2015) (Kazuyoshi et al., 2003) (Lin and Lin, 1999) (Wong et al., 2012)
(+)-Catechin (17)			

(-)-epi-catechin (18)	<i>T. theifer</i>	<i>Z. serrata</i>	(Tung-Hu et al., 2010)
(-)-epicatechin-3- <i>O</i> -gallate (19) [107]	<i>G. braunii</i>	<i>P. thonningii</i>	(Ja'afar et al., 2017)
(-)-epigallocatechin-3- <i>O</i> -gallate (20) [107]	<i>G. braunii</i>	<i>P. thonningii</i>	(Muhammad et al., 2022)
Quercetin 3- <i>O</i> - α -L-arabinoside (21)	<i>P. austroarabica</i>	NR	(Danladi et al., 2010)
Cyanidin 3- <i>O</i> - β -glucopyranoside (22)	<i>L. kaoi</i>	NR	(Lin and Lin, 1999)
Nicotiflorin (23)	<i>S. atropurpurea</i>	<i>T. sinensis</i>	(Kazuyoshi et al., 2003)
Hyperoside (24)	<i>S. atropurpurea</i>	<i>T. sinensis</i>	(Kazuyoshi et al., 2003)
Isoquercitrin (25)	<i>S. parasitica</i>	<i>N. indica</i> ,	(Quan-Yu et al., 2015)
(-)-catechin (26)	<i>P. capitata</i>	<i>C. spectabilis</i>	(Tung-Hu et al., 2010)
2',6'-dihydroxydihydrochalcone (27)	<i>T. buvumae</i>	<i>F. virosa</i>	(Roberts et al., 2011)
2',4',6'-trihydroxydihydrochalcone-4'- <i>O</i> - β -D-glucoside (28)	<i>T. constrictiflorus</i>	<i>E. abyssinica</i>	(Roberts et al., 2011)
Pinocembrin 7- <i>O</i> - β -D-glucoside (29)	<i>P. usuiensis</i>	<i>F. notalensis</i>	(Roberts et al., 2011)
Kaempferol 3- <i>O</i> - α -D-rhamnoside (30)	<i>T. acutifolius</i>	NR	(Soberón et al., 2010)
Kaempferol 3,7-di- <i>O</i> - β -D-glucoside (31)	<i>T. acutifolius</i>	NR	(Soberón et al., 2010)
AC trimer (32)	<i>L. kaoi</i>	NR	(Lin and Lin, 1999)
3- <i>O</i> -(3,4,5-trimethoxybenzoyl)-(-)-epicatechin (33)	<i>L. kaoi</i>	NR	(Lin and Lin, 1999)
(-)-epicatechin-3- <i>O</i> -(3''- <i>O</i> -methyl)-gallate (34)	<i>L. micranthus</i>	<i>H. brasiliensis</i>	(Matthias et al., 2014)
Peltatoside (35)	<i>L. micranthus</i>	<i>H. brasiliensis</i>	(Matthias et al., 2014)
-(-) catechin-7- <i>O</i> -rhamnoside (36)	<i>L. micranthus</i>	<i>K. acuminata</i>	(Omeje et al., 2012)
4'-methoxy catechin-7- <i>O</i> -rhamnoside (37)	<i>L. micranthus</i>	<i>K. acuminata</i>	(Omeje et al., 2012)
-(-) catechin-3- <i>O</i> -rhamnoside (38)	<i>L. micranthus</i>	<i>K. acuminata</i>	(Omeje et al., 2012)
3-methoxy quercetin (39)	<i>L. micranthus</i>	<i>K. acuminata</i>	(Edwin et al., 2014)
Catechin-4 - <i>O</i> -gallate (40)	<i>P. austroarabica</i>	NR	(Goda et al., 2022)
Pectolinarigenin (41)	<i>P. austroarabica</i>	NR	(Goda et al., 2022)
Dillenitin-3- <i>O</i> -glucoside (42)	<i>P. austroarabica</i>	NR	(Goda et al., 2022)
Rhamnetin 3- <i>O</i> - α -L-rhamnoside (43)	<i>L. tanakae</i>	<i>Quercus</i> and <i>Betula</i>	(Young-Kyoong et al., 2004)
Rhamnetin (44)	<i>G. braunii</i>	<i>T. catappa</i>	(Danladi et al., 2010)
Rhamnocrin 3- <i>O</i> - α -L-rhamnoside (45)	<i>G. braunii</i>	<i>T. catappa</i>	(Danladi et al., 2010)
Naringenin (46)	<i>L. tanakae</i>	<i>Quercus</i> and <i>Betula</i>	(Young-Kyoong et al., 2004)
Eriodictyol (47)	<i>V. album</i>	NR	(Yang et al., 2011)
Homoeriodictyol (48)	<i>V. articulatum</i>	NR	(Haizhen et al., 2015)
7- <i>O</i> - β -D-glucopyranoside (49)	<i>V. album</i>	NR	(Yang et al., 2011)
Homoeriodictyol 7- <i>O</i> - β -D-glucopyranoside (50)	<i>V. articulatum</i>	NR	(Haizhen et al., 2015)
3',4'- <i>O</i> -dimethyltaxifolin (51)	<i>V. colaratum</i>	NR	(Rui et al., 2012)
5,4'-dihydroxyflavanone 3,5,7,4'-tetrahydroxy-3'-methoxyflavanone (52)	<i>V. album</i>	NR	(Yang et al., 2011)
3,7,3'-tri- <i>O</i> -methylquercetin-4'- <i>O</i> - β -D-apiofuranosyl-(1 \rightarrow 2)- <i>O</i> - β -D-glucopyranoside (53)	<i>V. articulatum</i>	NR	(Yang et al., 2011)
7,3'-di- <i>O</i> -methylquercetin-4'- <i>O</i> - β -D-glucopyranosyl-3- <i>O</i> -[6''-(3-hydroxy-3-methylglutaroyl)]- α -D-glucopyranoside (54)	<i>V. album</i>	NR	(Nguyen et al., 2013)
7,3'-di- <i>O</i> -methylquercetin-4'- <i>O</i> - β -D-glucopyranosyl-3- <i>O</i> -[(6'''' \rightarrow 5''')- <i>O</i> -1''''-(sinap-4-yl)- β -D-glucopyranosyl-6''-(3-hydroxy-3-methylglutaroyl)]- α -D-glucopyranoside (55)	<i>V. album</i>	NR	(Nguyen et al., 2013)

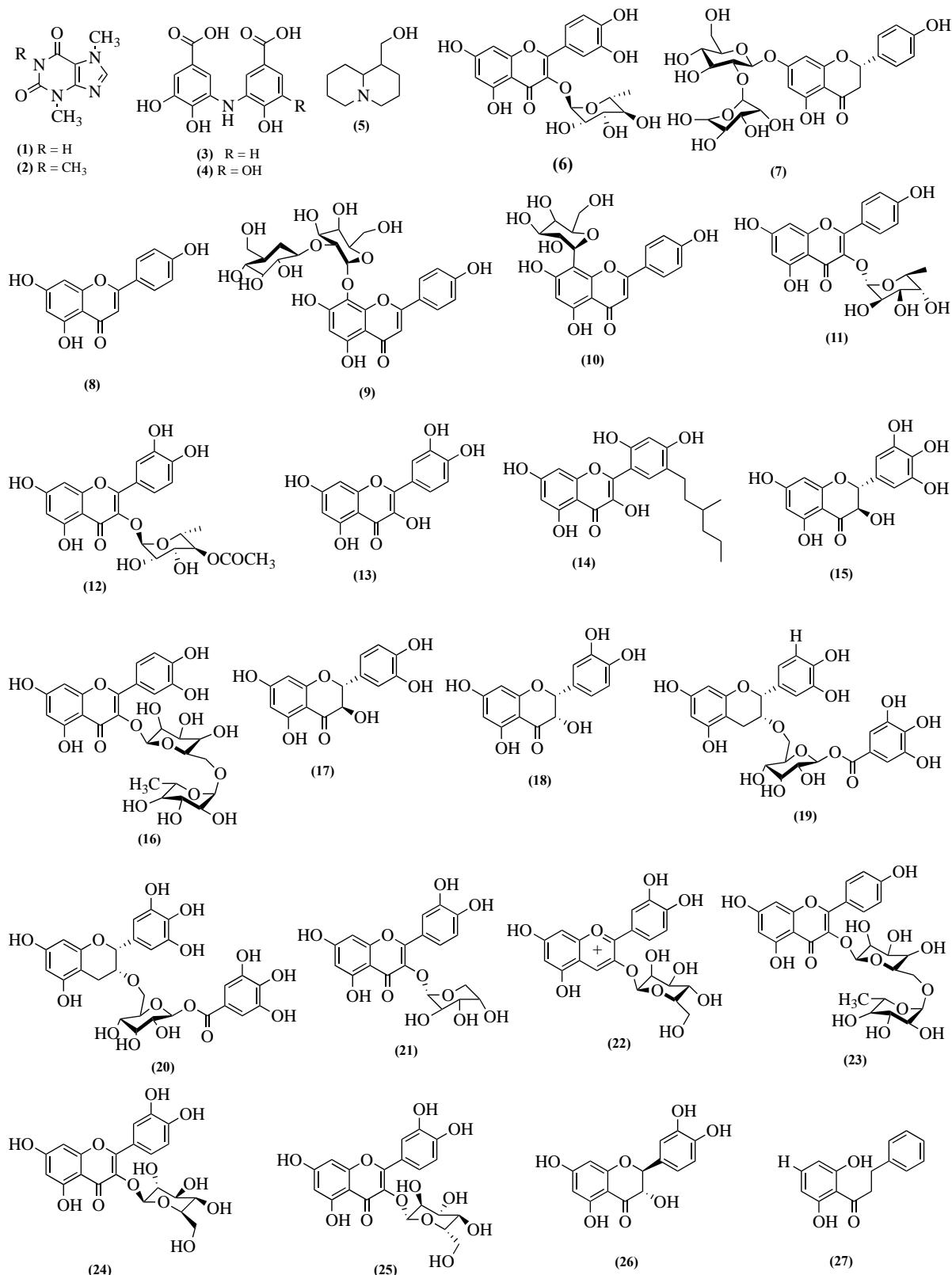
(2S)-5-hydroxy-7,3'-dimethoxyflavanone-4'-O- β -D-apiofuranosyl-(1 \rightarrow 5)-O- β -D-apiofuranosyl-(1 \rightarrow 2)-O- β -D-glucopyranoside (56)	<i>V. album</i>	NR	(Nguyen et al., 2013)
3'-methoxyapiin (57)	<i>V. album</i>	NR	(Nguyen et al., 2013)
Homoflavoyadorinin-B (58)	<i>V. album</i>	NR	(Nguyen et al., 2013)
(2S)-5-hydroxy-7,3'-dimethoxyflavanone-4'-O- β -[apiofuranosyl-(1 \rightarrow 2)]-glucopyranoside (59)	<i>V. album</i>	NR	(Nguyen et al., 2013)
(2S)-homoeriodictyol-7-O-[apiofuranosyl-(1 \rightarrow 2)]-glucopyranoside (60)	<i>V. album</i>	NR	(Nguyen et al., 2013)
Viscumneoside V (61)	<i>V. album</i>	NR	(Nguyen et al., 2013)
2R-Viscarticulide A (62)	<i>V. articulatum</i>	NR	(Haizhen et al., 2015)
2R-viscarticulide B (63)	<i>V. articulatum</i>	NR	(Haizhen et al., 2015)
2R-viscarticulide C (64)	<i>V. articulatum</i>	NR	(Haizhen et al., 2015)
2S-viscarticulide A (65)	<i>V. articulatum</i>	NR	(Haizhen et al., 2015)
2S-viscarticulide B (66)	<i>V. articulatum</i>	NR	(Haizhen et al., 2015)
2S-viscarticulide C (67)	<i>V. articulatum</i>	NR	(Haizhen et al., 2015)
Eriodictyol-7-O- β -D-glucoside (68)	<i>V. articulatum</i>	NR	(Haizhen et al., 2015)
Naringenin-7-O- β -D-glucoside (69)	<i>V. articulatum</i>	NR	(Haizhen et al., 2015)
Rhamnatin-3-O- β -D-apiosyl-(1 \rightarrow 2)- β -D-[6''-(3-hydroxy-3-methylglutaric methyl ester)]-glucoside (70)	<i>V. colaratum</i>	NR	(Rui et al., 2012)
Rhamnatin-3-O- β -D-apiosyl-(1 \rightarrow 2)- β -D-[6''-(3-hydroxy-3-methylglutarate)]-glucoside (71)	<i>V. colaratum</i>	NR	(Rui et al., 2012)
Isorhamnetin-3-O- β -D-glucoside (72)	<i>V. colaratum</i>	NR	(Rui et al., 2012)
Homoeriodictyol-7-O- β -D-apiosyl-(1 \rightarrow 5)- β -D-apiosyl-(1 \rightarrow 2)- β -D-glucoside (73)	<i>V. colaratum</i>	NR	(Rui et al., 2012)
(2S)-homoeriodictyol 7,4'-di-O- β -D-glucopyranoside (74)	<i>V. colaratum</i>	NR	(Hui et al., 2006)
(2R)-eriodictyol 7,4'-di-O- β -D-glucopyranoside (75)	<i>V. colaratum</i>	NR	(Hui et al., 2006)
Homoeriodictyol-7-O- β -D-[6-(3-hydroxybutanoyl) glucopyranoside] (76)	<i>V. colaratum</i>	NR	(Na et al., 2011)
Homoeriodictyol-7-O- β -D-[6-(3-hydroxybutanoyl) glucopyranosyl](1 \rightarrow 2)- β -D-glucopyranoside (viscumneoside X) (77)	<i>V. colaratum</i>	NR	(Na et al., 2011)
Viscumneoside I (78)	<i>V. colaratum</i>	NR	(Na et al., 2011)
Viscumneoside III (79)	<i>T. sutchuenensis</i>	NR	(Liyuan et al., 2017)
Viscumneoside XII (80)	<i>V. album</i>	NR	(Jia-Kun et al., 2019)
Viscumneoside XIII (81)	<i>V. album</i>	NR	(Jia-Kun et al., 2019)
Viscumneoside XIV (82)	<i>V. colaratum</i>	NR	(Jia-Kun et al., 2019)
4',5-dihydroxy-3'-methoxy-7-(2-O- α -L-rhamnopyranosyl- β -D-glucopyranosyloxy) flavanone (83)	<i>V. album</i>	NR	(Jia-Kun et al., 2019)
Catechin-5-O-(6-O-galloyl- β -glucopyranoside) (84)	<i>T. theifer</i>	<i>Z. serrata</i>	(Tung-Hu et al., 2010)
Quercetin-3-O-(6-O-galloyl- β -glucopyranoside) (85)	<i>T. theifer</i>	<i>Z. serrata</i>	(Tung-Hu et al., 2010)
Quercetin-3-O- β -glucuronide (86)	<i>T. theifer</i>	<i>Z. serrata</i>	(Tung-Hu et al., 2010)
Quercetin-3-O- β -glucuronic acid methyl ester (87)	<i>T. theifer</i>	<i>Z. serrata</i>	(Tung-Hu et al., 2010)
Quercetin-3-O- β -glucuronic acid butyl ester (88)	<i>T. theifer</i>	<i>Z. serrata</i>	(Tung-Hu et al., 2010)
Kaempferol-3-O- β -glucopyranoside (89)	<i>T. theifer</i>	<i>Z. serrata</i>	(Tung-Hu et al., 2010)
Quercetin-4'-O- β -glucopyranoside (90)	<i>T. theifer</i>	<i>Z. serrata</i>	(Tung-Hu et al., 2010)
Iisosakuranetin (91)	<i>T. sutchuenensis</i>	NR	(Liyuan et al., 2017)
Sakuranetin (92)	<i>V. album</i>	NR	(Nonato de et al., 2018)
Naringenin-5-methyl-ether (93)	<i>V. album</i>	NR	(Nonato de et al., 2018)
Quercetin 3,3',4'-trimethyl ether (94)	<i>T. sutchuenensis</i>	NR	(Liyuan et al., 2017)
Kaempferol-3,7-bisrhamnoside (95)	<i>T. sutchuenensis</i>	NR	(Liyuan et al., 2017)
Avicularin (96)	<i>G. braunii</i>	<i>P. thonningii</i>	(Ja'afar et al., 2017)
7-hydroxy-4',5,6-trimethoxyflavone (97)	<i>D. falcata</i>	NR	(Muhammad et al., 2022)
2',4',6-trimethoxyflavone (98)	<i>L. acutifolius</i>	NR	(Kumar et al., 2022)
3',4',5-trihydroxy-6,7,8-trimethoxyflavone (99)	<i>T. acutifolius</i>	<i>T. tacaca</i>	(Ticona et al., 2020)
	<i>L. acutifolius</i>	NR	(Apaza et al., 2019)
	<i>T. acutifolius</i>	<i>T. tacaca</i>	(Ticona et al., 2020)
	<i>T. acutifolius</i>	NR	(Apaza et al., 2019)

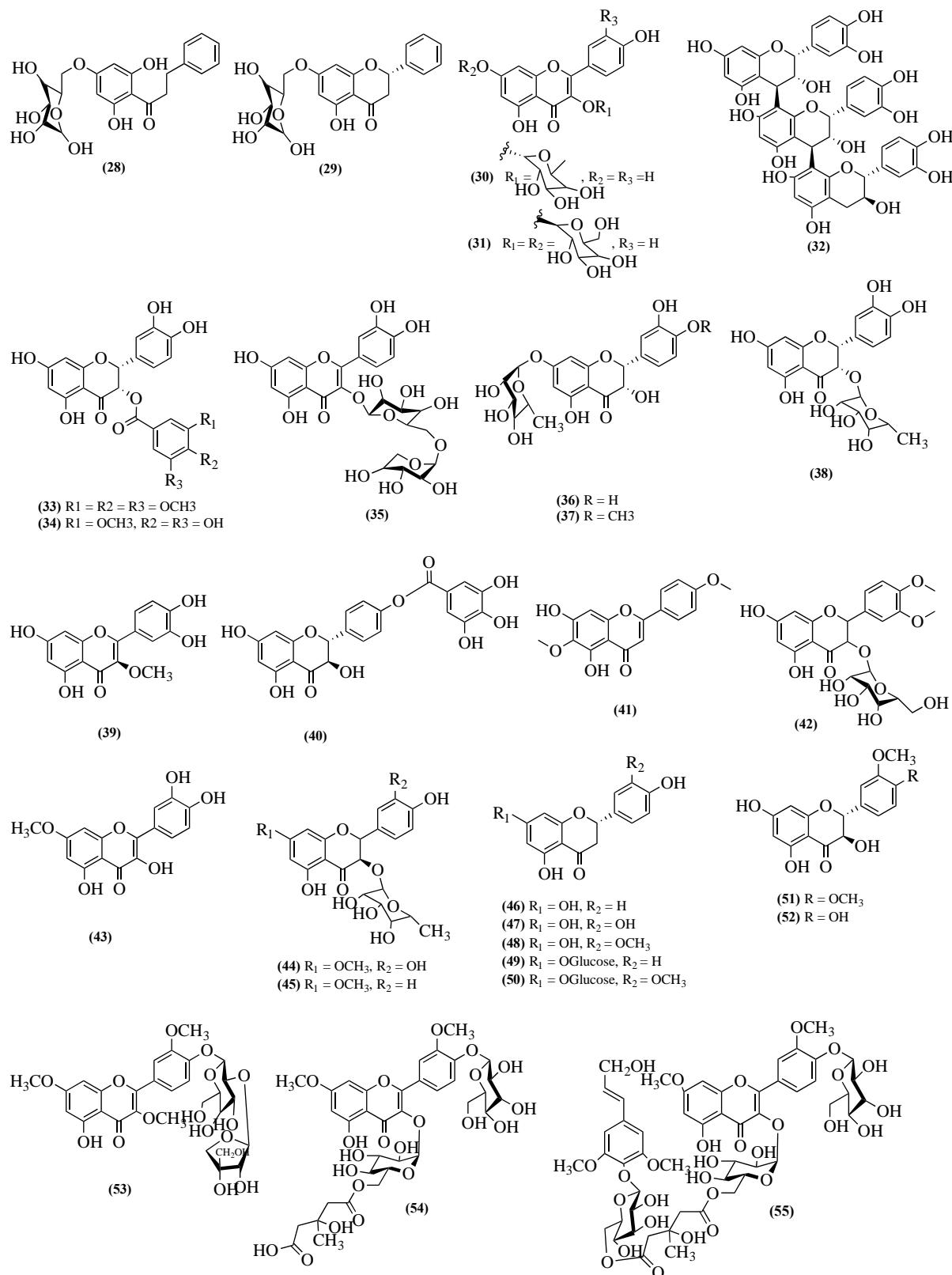
2'4'-dihydroxy-6'-methoxy-chalcone (100)	<i>L. acutifolius</i>	NR	(Ticona et al., 2020)
4',5-dihydroxy-6,7,8-trimethoxyflavone (101)	<i>T. acutifolius</i>	<i>T. tacaca</i>	(Apaza et al., 2019)
Quercetin 4'-methyl ether (102)	<i>L. acutifolius</i>	NR	(Ticona et al., 2020)
4'-methoxy-3',5,7-trihydroxyflavone (103)	<i>T. acutifolius</i>	<i>T. tacaca</i>	(Apaza et al., 2019)
Quercetin-3-O-rhamnoside (104)	<i>T. pentagonia</i>	Avocado tree	(Ikome et al., 2020)
Quercetin 3-O-rhamnoside 4'-methyl ether (105)	<i>T. pentagonia</i>	Avocado tree	(Ikome et al., 2020)
5-dimethoxy-7-hydroxy 8-dimethoxy flavanone (106)	<i>T. pentagonia</i>	Avocado tree	(Ikome et al., 2020)
4,5-dimethoxy-4'-hydroxy flavanone (107)	<i>V. album</i>	<i>J. regia</i>	(Maher et al., 2021)
5,7-di-methoxy-4-O-β-D-glucopyranoside flavanone (108)	<i>V. album</i>	<i>J. regia</i>	(Maher et al., 2021)
5-methoxy-7-O-β-D-glucopyranoside flavanone (109)	<i>V. album</i>	<i>J. regia</i>	(Maher et al., 2021)
PHENOLIC COMPOUNDS			
(1R,5S,7S)-7-[2-(4-hydroxyphenyl)ethyl]-2,6-dioxabicyclo[3.3.1]nonan-3-one (110)	<i>G. dinklagei</i>	<i>M. esculenta</i>	(Mkounga et al., 2016)
4-hydroxy-3,5-dimethoxy benzoic acid (111)	<i>P. capitata</i>	<i>C. spectabilis</i>	(Bruno et al., 2015)
Gallic acid (112)	<i>T. bangwensi</i>	<i>C. occidentalis</i>	(Harmine et al., 2017)
Methyl gallate (113)	<i>G. braunii</i>	<i>P. thonningii</i>	(Ja'afar et al., 2017)
Methyl syringate (114)	<i>G. braunii</i>	<i>P. thonningii</i>	(Muhammad et al., 2022)
Methyl 3,4-O-dimethyl gallate (115)	<i>T. globiferus</i>	<i>P. biglobosa</i>	(Anyam et al., 2022)
3,4,5-trimethoxy methyl benzoate (116)	<i>G. dinklagei</i>	<i>M. esculenta</i>	(Mkounga et al., 2016)
3-hydroxy-4,5-dimethoxy methyl benzoate (117)	<i>D. falcate</i>	<i>S. robusta</i>	(Mallavadhani et al., 2006)
Dodoneine (118)	<i>P. austroarabica</i>	NR	(Goda et al., 2022)
4,6-Dihydroxy-8-[2-(4-hydroxy-phenyl)-ethyl]-oxocan-2-one (119)	<i>V. album</i>	<i>J. regia</i>	(Maher et al., 2021)
6-Hydroxy-8-[2-(4-hydroxy-phenyl)-ethyl]-5,6,7,8-tetrahydro-oxocin-2-one (120)	<i>T. bangwensis</i>	NR	(Goda et al., 2022)
Tripodanthoside (121)	<i>T. bangwensis</i>	NR	(Patrick-Iwuanyanwu et al., 2014)
methyl 2,6-dihydroxy-4-methoxybenzoate (122)	<i>T. dodoneifolius</i>	<i>V. paradoxa</i>	(Patrick-Iwuanyanwu et al., 2014)
methyl 3,5-dihydroxy-4-methoxybenzoate (123)	<i>G. braunii</i>	<i>P. thonningii</i>	(Maurice et al., 2007)
methyl 3-methyl-4-hydroxybenzoate (124)	<i>G. braunii</i>	<i>P. thonningii</i>	(Ja'afar et al., 2017)
Guaiacol (125)	<i>T. globiferus</i>	<i>P. thonningii</i>	(Muhammad et al., 2022)
4-formaldehyde phenone (126)	<i>G. braunii</i>	<i>P. biglobosa</i>	(Anyam et al., 2022)
6-methoxy-2H-inden-5-ol (127)	<i>G. braunii</i>	<i>P. thonningii</i>	(Abdullahi et al., 2014)
rel-(1R,5S,7S)-7-[2-(4-O-galloylphenyl)ethyl]-2,6-dioxabicyclo-[3.3.1] nonan-3-one (128)	<i>G. braunii</i>	<i>P. thonningii</i>	(Abdullahi et al., 2014)
p-hydroxyphenylacetic acid (129)	<i>T. acutifolius</i>	NR	(Soberón et al., 2010)
5,7-dihydroxychromone (130)	<i>G. braunii</i>	<i>L. leucocephala</i>	(Ayoola et al., 2020)
Centrolobol (131)	<i>G. braunii</i>	<i>L. leucocephala</i>	(Oriola et al., 2021)
Acerogenin G (132)	<i>G. braunii</i>	<i>L. leucocephala</i>	(Oriola et al., 2021)
(3S,5R)-3-hydroxy-5-methoxy-1,7-bis(4-hydroxyphenyl)-6E-heptene (133)	<i>G. braunii</i>	<i>L. leucocephala</i>	(Oriola et al., 2021)
(3S,5S)-3-hydroxy-5-methoxy-1,7-bis(4-hydroxyphenyl)-6E-heptene (134)	<i>V. album</i>	NR	(Oriola et al., 2021)
(3S)-3-hydroxy-1,7-bis(4-hydroxyphenyl)-6E-hepten-5-one (135)	<i>V. album</i>	NR	(Oriola et al., 2021)
1,7-di-(3',4'-dihydroxyphenyl)-4-hepten-3-one, hirsutanone (136)	<i>T. sutchuenensis</i>	NR	(Nguyen et al., 2013)
Taxilluside A (137)	<i>V. cruciatum</i>	<i>P. amygdalus</i>	(Liyuan et al., 2017)
	<i>T. chinensis</i>	NR	(Carmen et al., 2001)
			(Bo et al., 2013)

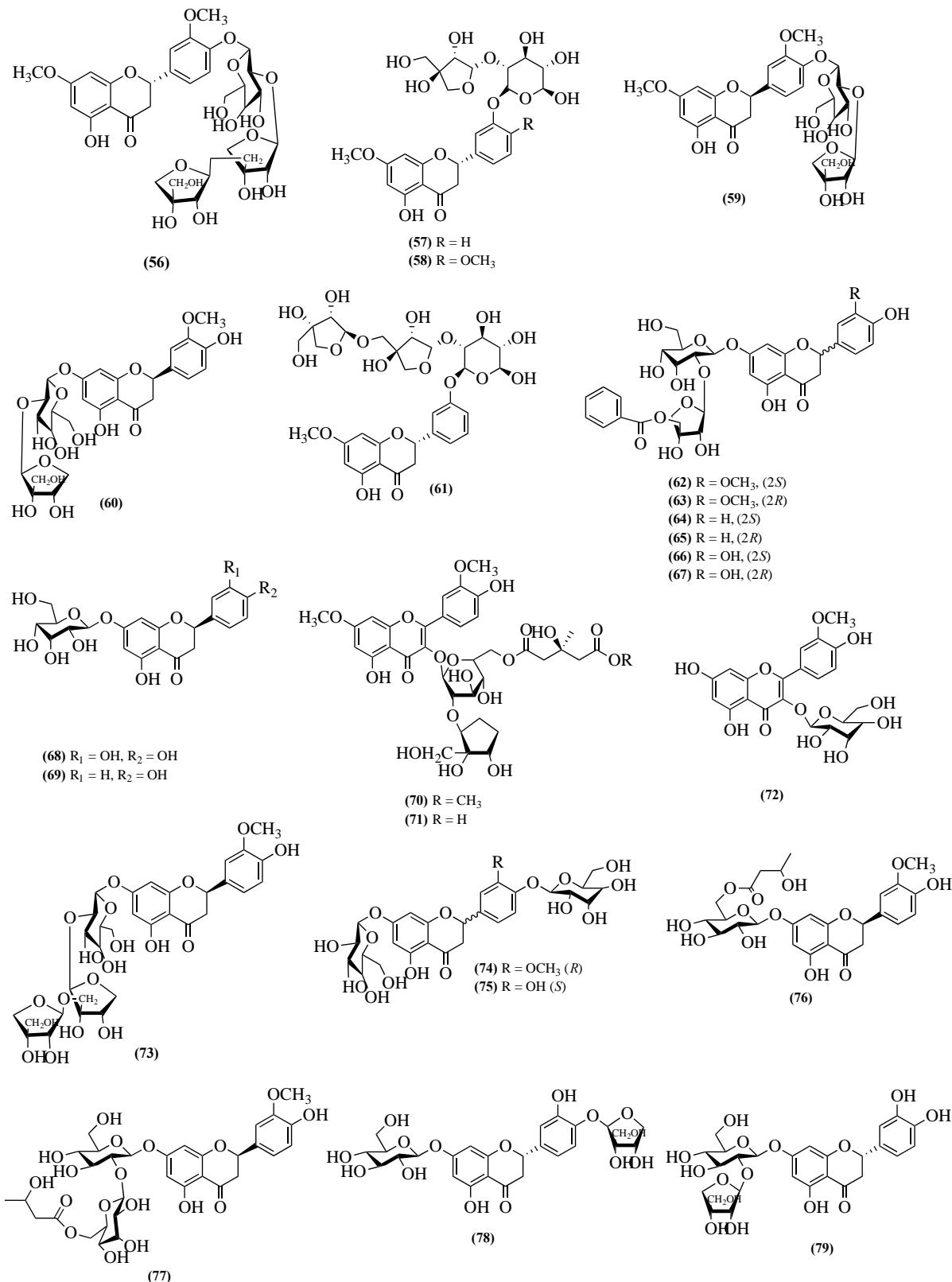
Taxilluside B (138)	<i>T. chinensis</i>	NR	(Bo et al., 2013)
Taxilluside C (139)	<i>T. chinensis</i>	NR	(Bo et al., 2013)
Taxilluside D (140)	<i>T. chinensis</i>	NR	(Bo et al., 2013)
(+)-hannokinol (141)	<i>T. sutchuenensis</i>	NR	(Liyuan et al., 2017)
Meso-hannokinol (142)	<i>T. sutchuenensis</i>	NR	(Liyuan et al., 2017)
2,3-dihydro-4-hydroxy-3,6,9-trimethylnaphtho[1,8-bc]pyran-7,8-dione (143)	<i>D. falcata</i>	NR	(Kumar et al., 2022)
4-hydroxy-3-methoxybenzoic acid (144)	<i>D. falcata</i>	NR	(Kumar et al., 2022)
Emodin (145)	<i>P. austroarabica</i>	NR	(Goda et al., 2022)
emodin-8-O-glucoside (146)	<i>P. austroarabica</i>	NR	(Goda et al., 2022)
Chrysophanic acid (147)	<i>P. austroarabica</i>	NR	(Goda et al., 2022)
Chrysophanic acid-8-O-glucoside (148)	<i>P. austroarabica</i>	NR	(Goda et al., 2022)
3-(4-hydroxy-3,5-dimethoxy)-phenyl-2-propenylβ-D-glucopyranoside (149)	<i>V. album</i>	<i>J. regia</i>	(Maher et al., 2021)
(7) 3-(4-hydroxy-3,5-dimethoxy)-phenyl-2-epropenol (150)	<i>V. album</i>	<i>J. regia</i>	(Maher et al., 2021)
Caffeic acid (151)	<i>V. album</i>	NR	(Nonato de et al., 2018)
Chlorogenic acid (152)	<i>V. album</i>	NR	(Nonato de et al., 2018)
syringenin 4-O-glucoside (153)	<i>V. album</i>	NR	(Nonato de et al., 2018)
syringenin 4-O-apiosyl glucoside (154)	<i>V. album</i>	NR	(Nonato de et al., 2018)
alangilignoside C and ligalbumoside A (155)	<i>V. album</i>	NR	(Nonato de et al., 2018)
LIGNANS AND NEOLIGNANS			
Aviculin (156)	<i>S. atropurpurea</i>	<i>T. sinensis</i>	(Kazuyoshi et al., 2003)
(1S, 3aR, 4S, 6aR)-1-[(3, 5-dimethoxyl- 4-hydroxyl) phenyl]-4-[(3-methoxyl-4-β-D-O-glucopyranosyl-5-hydroxyl) phenyl] -tetrahydro-1H, 3H-furo [3, 4-c] furan (viscoloratin) (157)	<i>V. colaratum</i>	NR	(Jun et al., 2007)
(+)-medioresinol (158)	<i>V. album</i>	NR	(Nguyen et al., 2013)
(+)-pinoresinol (159)	<i>V. album</i>	NR	(Nguyen et al., 2013)
<i>V. album</i>	<i>V. album</i>	NR	(Yang et al., 2011)
(-)lyoniresinol 3α-O-β-D-glucopyranoside (160)	<i>V. album</i>	NR	(Roberts et al., 2011)
(+)-lyoniresinol 3α-O-β-D-glucopyranoside (161)	<i>V. album</i>	NR	(Roberts et al., 2011)
Syringin (162)	<i>V. album</i>	NR	(Roberts et al., 2011)
Syringenin 4'-O-β-D-apiofuranosyl-(1→2)-β-O-D-glucopyranoside (163)	<i>V. album</i>	NR	(Roberts et al., 2011)
Syringaresinol (164)	<i>V. album</i>	NR	(Yang et al., 2011)
Syringaresinal-4'-O-β-D-glucopyranoside (165)	<i>V. album</i>	NR	(Yang et al., 2011)
Curulignan D (166)	<i>V. album</i>	NR	(Yang et al., 2011)
<i>threo</i> -(7 <i>R</i> , 8 <i>R</i>)-7-acetoxy-3',4'-dimethoxy-3,4-dimethoxy-D-8'-O,6'-neolignan (167)	<i>T. theifer</i>	NR	(Lie-Chwen et al., 2011)
<i>threo</i> -7-acetoxy-3'-methoxy-3,4-dimethoxy-D-7'-8,O,4'-neolignan (168)	<i>T. theifer</i>	NR	(Lie-Chwen et al., 2011)
7 <i>R</i> , 8 <i>R</i> , 3'(<i>R</i>)-7-acetoxy-3',4'-dimethoxy-3,4-dimethoxy-6'-oxo-D-1',4',8'-3' lignin (169)	<i>T. theifer</i>	NR	(Lie-Chwen et al., 2011)
Tremulacin (170)	<i>T. sutchuenensis</i>	NR	(Liyuan et al., 2017)
TRITERPENES			
13,27-cycloursane (171)	<i>G. braunii</i>	<i>L. leucocephala</i>	(Ayoola et al., 2020)
Phyllanthone (172)	<i>G. braunii</i>	<i>L. leucocephala</i>	(Oriola et al., 2021)
Globraunone (173)	<i>G. braunii</i>	<i>L. leucocephala</i>	(Oriola et al., 2021)
Globraunine A (174)	<i>G. braunii</i>	<i>P. thonningii</i>	(Muhammad et al., 2020)
Globraunine B (175)	<i>G. braunii</i>	<i>P. thonningii</i>	(Muhammad et al., 2020)
Globraunine C (176)	<i>G. braunii</i>	<i>P. thonningii</i>	(Muhammad et al., 2020)
Globraunine D (177)	<i>G. braunii</i>	<i>P. thonningii</i>	(Muhammad et al., 2020)
Globraunine E (178)	<i>G. braunii</i>	<i>P. thonningii</i>	(Muhammad et al., 2020)
Globraunine F (179)	<i>G. braunii</i>	<i>P. thonningii</i>	(Muhammad et al., 2020)
Globimetulin A (180)	<i>G. dinklagei</i>	<i>M. esculenta</i>	(Mkounga et al., 2016)
Globimetulin B (181)	<i>G. dinklagei</i>	<i>M. esculenta</i>	(Mkounga et al., 2016)
Globimetulin C (182)	<i>G. dinklagei</i>	<i>M. esculenta</i>	(Mkounga et al., 2016)
Friedelin (183)	<i>S. parasitica</i>	<i>N. indicum</i>	(Quan-Yu et al., 2015)
	<i>G. dinklagei</i>	<i>M. esculenta</i>	(Mkounga et al., 2016)

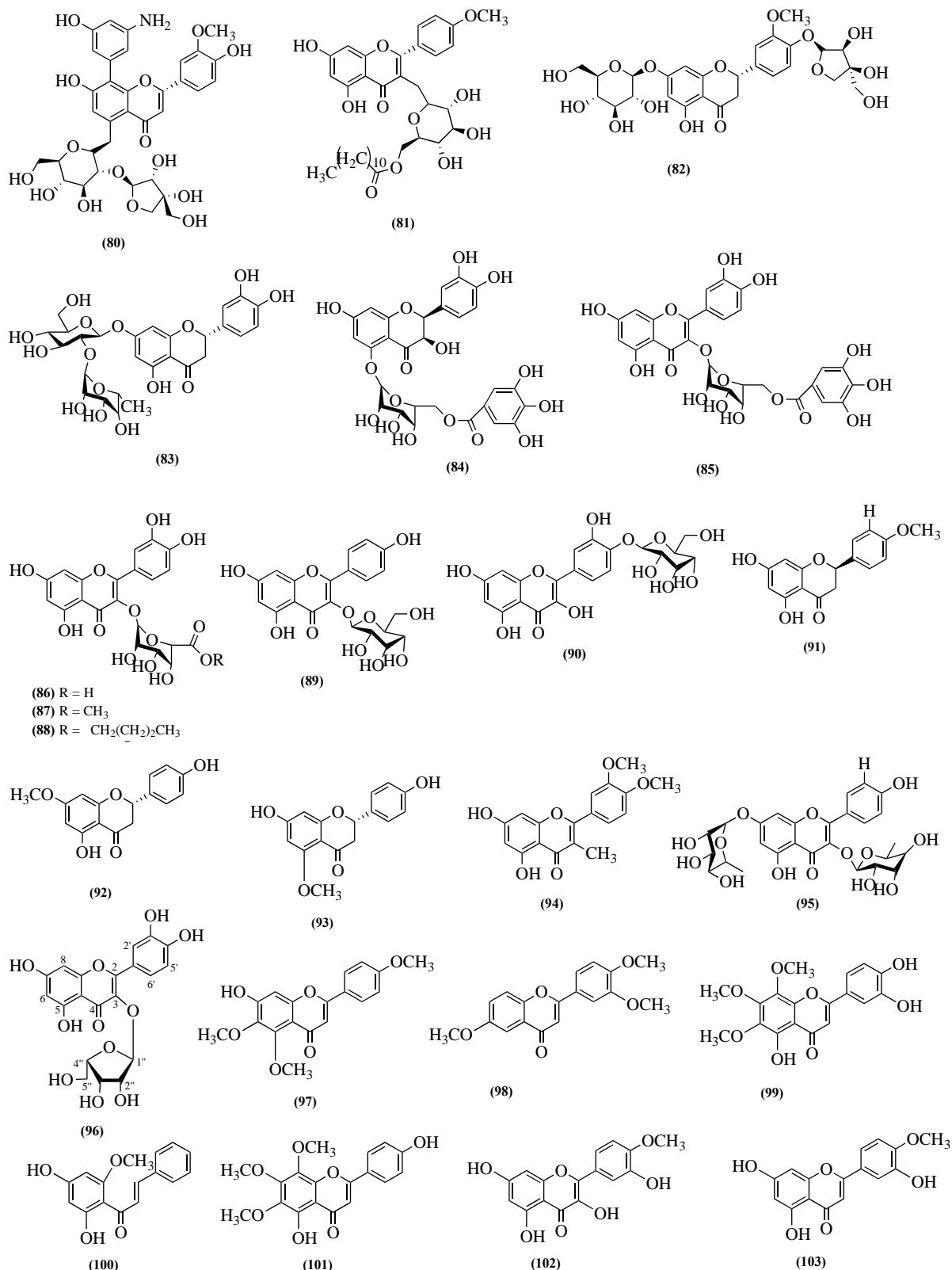
Betulin (216)	<i>P. capitata</i>	<i>C. spectabilis</i>	(Bruno et al., 2015)
Betulinic acid (217)	<i>V. album</i>	NR	(Yang et al., 2011)
	<i>V. album</i>	NR	(Nguyen et al., 2013)
	<i>L. acacie</i>	NR	(Noman et al., 2020)
	<i>T. sessilifolius</i>	<i>P. guajava</i>	(Tarfà et al., 2022)
Bangwaoleanenes A (218)	<i>T. bangwensis</i>	<i>C. occidentalis</i>	(Harmine et al., 2017)
Bangwaoleanenes B (219)	<i>T. bangwensis</i>	<i>C. occidentalis</i>	(Harmine et al., 2017)
Bangwaoleanenes C (220)	<i>T. bangwensis</i>	<i>C. occidentalis</i>	(Harmine et al., 2017)
Bangwaoleanenes D (221)	<i>T. bangwensis</i>	<i>C. occidentalis</i>	(Harmine et al., 2017)
Bangwaoleanenes E (222)	<i>T. bangwensis</i>	<i>C. occidentalis</i>	(Harmine et al., 2017)
Bangwaursenes A (223)	<i>T. bangwensis</i>	<i>C. occidentalis</i>	(Harmine et al., 2017)
Bangwaursenes B (224)	<i>T. bangwensis</i>	<i>C. occidentalis</i>	(Harmine et al., 2017)
3 β -acetoxy-urs-12,13-ene-11-one (225)	<i>T. bangwensis</i>	<i>C. occidentalis</i>	(Harmine et al., 2017)
3 β -acetoxy-11 α -hydroxyurs-12,13-ene (226)	<i>T. bangwensis</i>	<i>C. occidentalis</i>	(Harmine et al., 2017)
11 α ,12 α - oxidotaraxeryl acetate (227)	<i>T. bangwensis</i>	<i>C. occidentalis</i>	(Harmine et al., 2017)
β -amyrin acetate (228)	<i>T. bangwensis</i>	<i>C. occidentalis</i>	(Harmine et al., 2017)
7 β ,15 α -dihydroxyl-lup-20(29)-ene-3 β -stearate (229)	<i>V. album</i>	NR	(Nguyen et al., 2013)
7 β ,15 α -dihydroxyl-lup-20(29)-ene-3 β -eicosanoate (230)	<i>L. micranthus</i>	<i>K. acuminata</i>	(Ogechukwu et al., 2011)
Stigmast-7, 20(21)-diene-3 β -hydroxy-6-one (231)	<i>L. micranthus</i>	<i>K. acuminata</i>	(Ogechukwu et al., 2011)
3 β -hydroxy-stigmast-23-ene (232)	<i>L. micranthus</i>	<i>K. acuminata</i>	(Ogechukwu et al., 2011)
Erythrodiol (233)	<i>V. album</i>	NR	(Nguyen et al., 2013)
Oleanolic acid (234)	<i>V. album</i>	NR	(Yang et al., 2011)
Stigmast-4-en-3-one (235)	<i>T. sutchuenensis</i>	NR	(Liyuan et al., 2017)
Stigmast-5-en-3 β -ol (236)	<i>D. falcata</i>	NR	(Kumar et al., 2022)
3 β -hydroxylup-20(29)-en-28-oic acid (237)	<i>D. falcata</i>	NR	(Kumar et al., 2022)
3 β -hydroxyolean-12-en-28-oic acid (238)	<i>D. falcata</i>	NR	(Kumar et al., 2022)
OTHER GROUP OF COMPOUNDS			
(Z)-9-octadecenoic acid (Oleic acid) (239)	<i>S. atropurpurea</i>	<i>T. sinensis</i>	(Kazuyoshi et al., 2003)
(Z,Z)-octadeca-9,12-dienoic acid (Linoleic acid) (240)	<i>S. atropurpurea</i>	<i>T. sinensis</i>	(Kazuyoshi et al., 2003)
(Z,Z,Z)-octadeca-9,12,15-trienoic acid (Linolenic acid) (241)	<i>S. atropurpurea</i>	<i>T. sinensis</i>	(Kazuyoshi et al., 2003)
Octadeca-8,10-diynoic acid (242)	<i>S. atropurpurea</i>	<i>T. sinensis</i>	(Kazuyoshi et al., 2003)
Octadec-12-ene-8,10-diynoic acid (243)	<i>S. atropurpurea</i>	<i>T. sinensis</i>	(Kazuyoshi et al., 2003)
Octadeca-8,10,12-triynoic acid (244)	<i>S. atropurpurea</i>	<i>T. sinensis</i>	(Kazuyoshi et al., 2003)
Icariside (245)	<i>S. atropurpurea</i>	<i>T. sinensis</i>	(Kazuyoshi et al., 2003)
Gitoxigenin 3-O- α -L-rhamnoside (246)	<i>S. parasitica</i>	<i>N. indicum</i>	(Quan-Yu et al., 2015)
Digitoxigenin 3-O- α -L-rhamnoside (247)	<i>S. parasitica</i>	<i>N. indicum</i>	(Quan-Yu et al., 2015)
Gitoxigenin 3-O- α -D-glucoside (248)	<i>S. parasitica</i>	<i>N. indicum</i>	(Quan-Yu et al., 2015)
Digitoxin 3-O- α -D-glucose (249)	<i>S. parasitica</i>	<i>N. indicum</i>	(Quan-Yu et al., 2016)
Behenic acid (250)	<i>S. parasitica</i>	<i>N. indicum</i>	(Quan-Yu et al., 2016)
Octacosyl alcohol (251)	<i>S. parasitica</i>	<i>N. indicum</i>	(Quan-Yu et al., 2016)
Perseitol (252)	<i>S. fusca</i>	<i>F. riedelii</i>	(Takashi et al., 2001)
1-desoxyribosa (253)	<i>T. bangwensis</i>	<i>C. occidentalis</i>	(Harmine et al., 2017)
Myo-inositol (254)	<i>T. bangwensis</i>	<i>C. occidentalis</i>	(Harmine et al., 2017)
Sorbitol (255)	<i>T. bangwensis</i>	<i>C. occidentalis</i>	(Harmine et al., 2017)
2,3-dimethoxy-benzo[a, b] cyclopentenyl-3',3',5'-trimethyl pyran-4-carboxylic acid (256)	<i>L.micranthus</i>	<i>K. acuminata</i>	(Edwin et al., 2011)
3,4,5-trimethoxy benzoate (257)	<i>L.micranthus</i>	<i>K. acuminata</i>	(Edwin et al., 2014)
Trans-phytol (258)	<i>V. album</i>	NR	(Nguyen et al., 2013)
Nerolidol (259)	<i>V. album</i>	NR	(Nguyen et al., 2013)
Nonadecan-1-ol (260)	<i>D. falcata</i>	NR	(Kumar et al., 2022)
Di-iso-octylphthalate (261)	<i>D. falcata</i>	NR	(Kumar et al., 2022)
2,6-Dimethylocta-2,7-diene-1,6-diol 6-O-[6'-O- β -D-apiofuranosyl]- β -D-glucopyranoside (262)	<i>V. album</i>	<i>A. vulgaris</i>	(Deliorman et al., 2001)
Octacosanoic acid (263)	<i>G. braunii</i>	<i>P. thonningii</i>	(Muhammad et al., 2022)
Octacosane (264)	<i>G. braunii</i>	<i>P. thonningii</i>	(Muhammad et al., 2020)
Octadecane (265)	<i>S. parasitica</i>	<i>P. pinnata</i>	(Muhammad et al., 2019)
Eicosane (266)	<i>S. parasitica</i>	<i>P. pinnata</i>	(Muhammad et al., 2019)

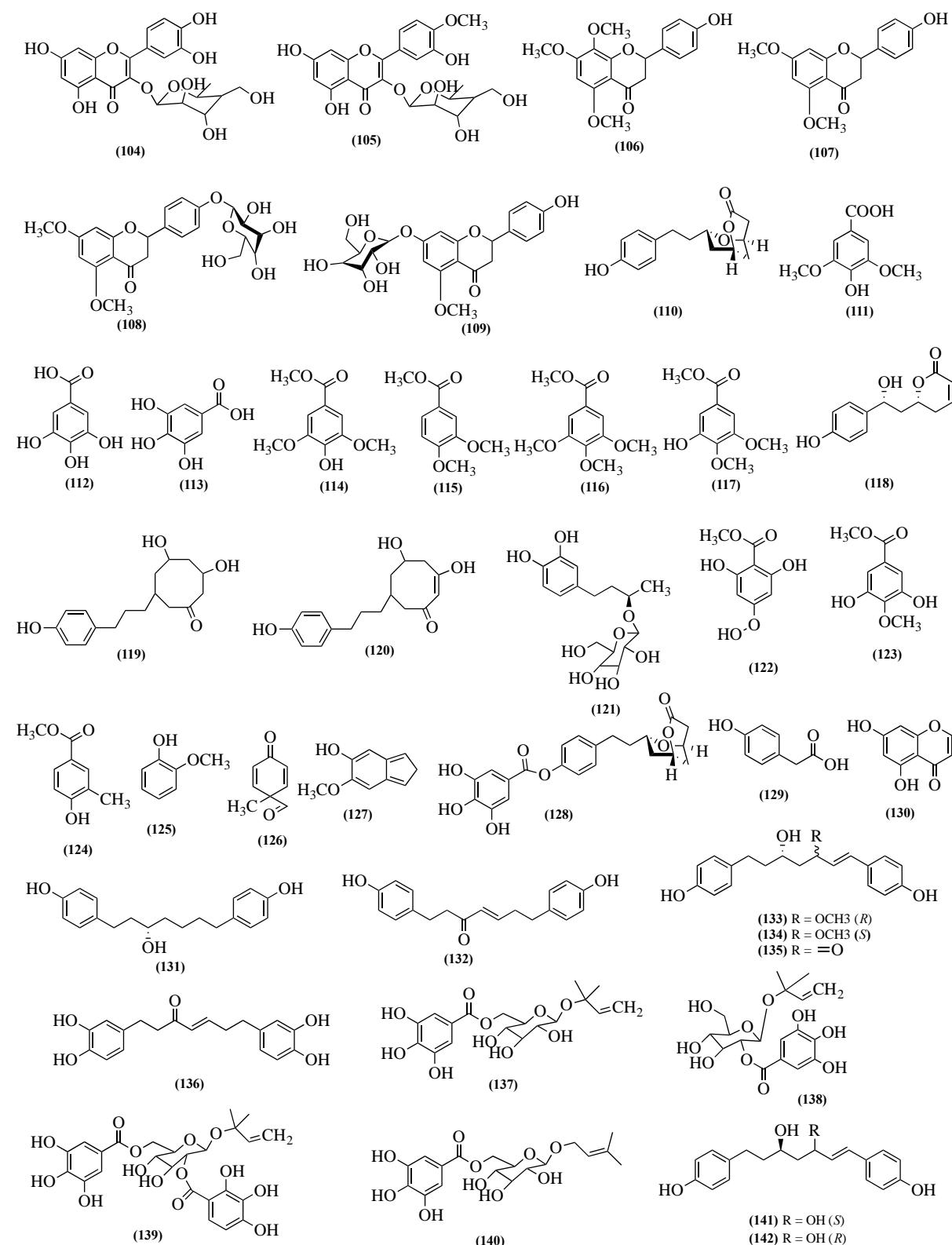
*NR = Not Reported

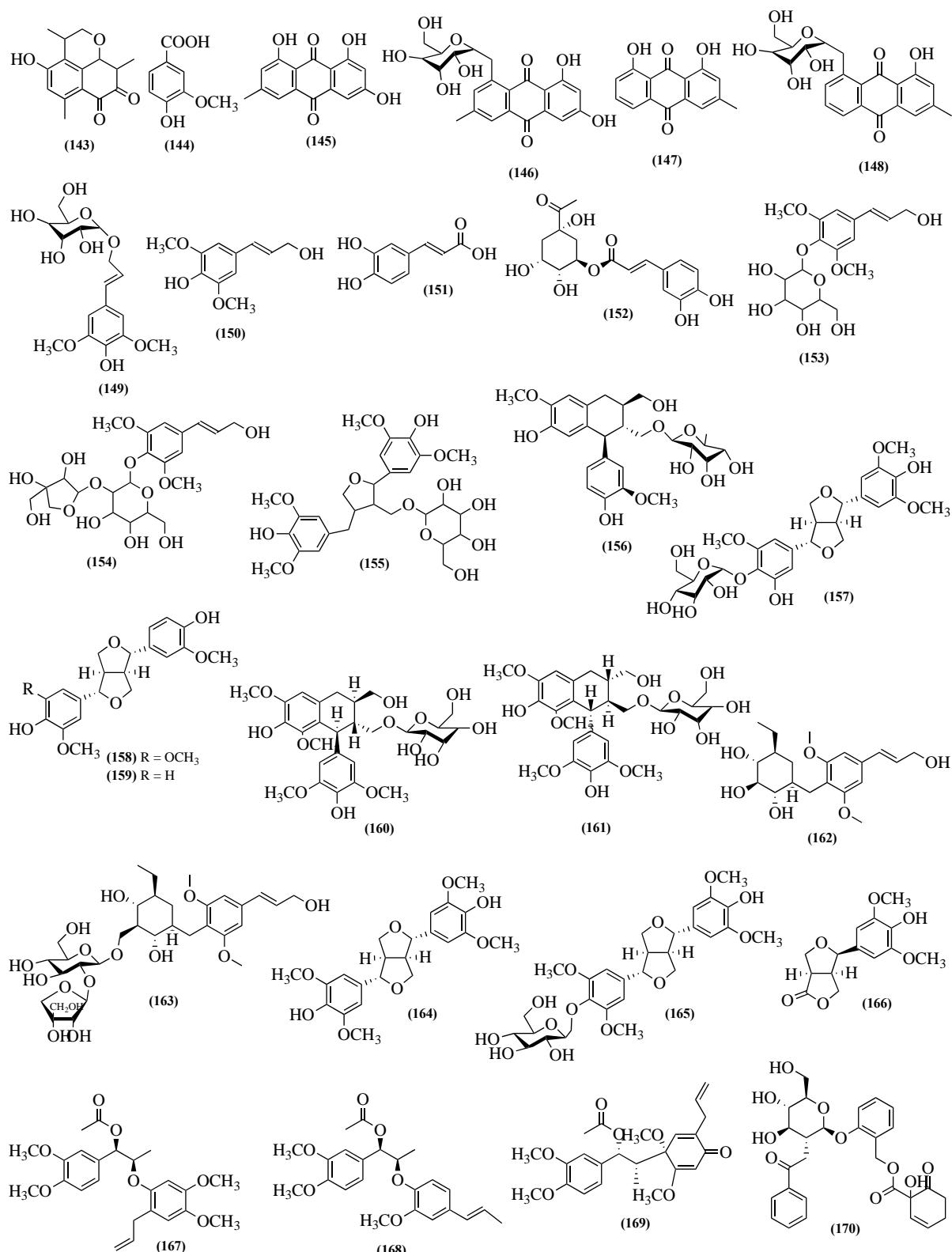


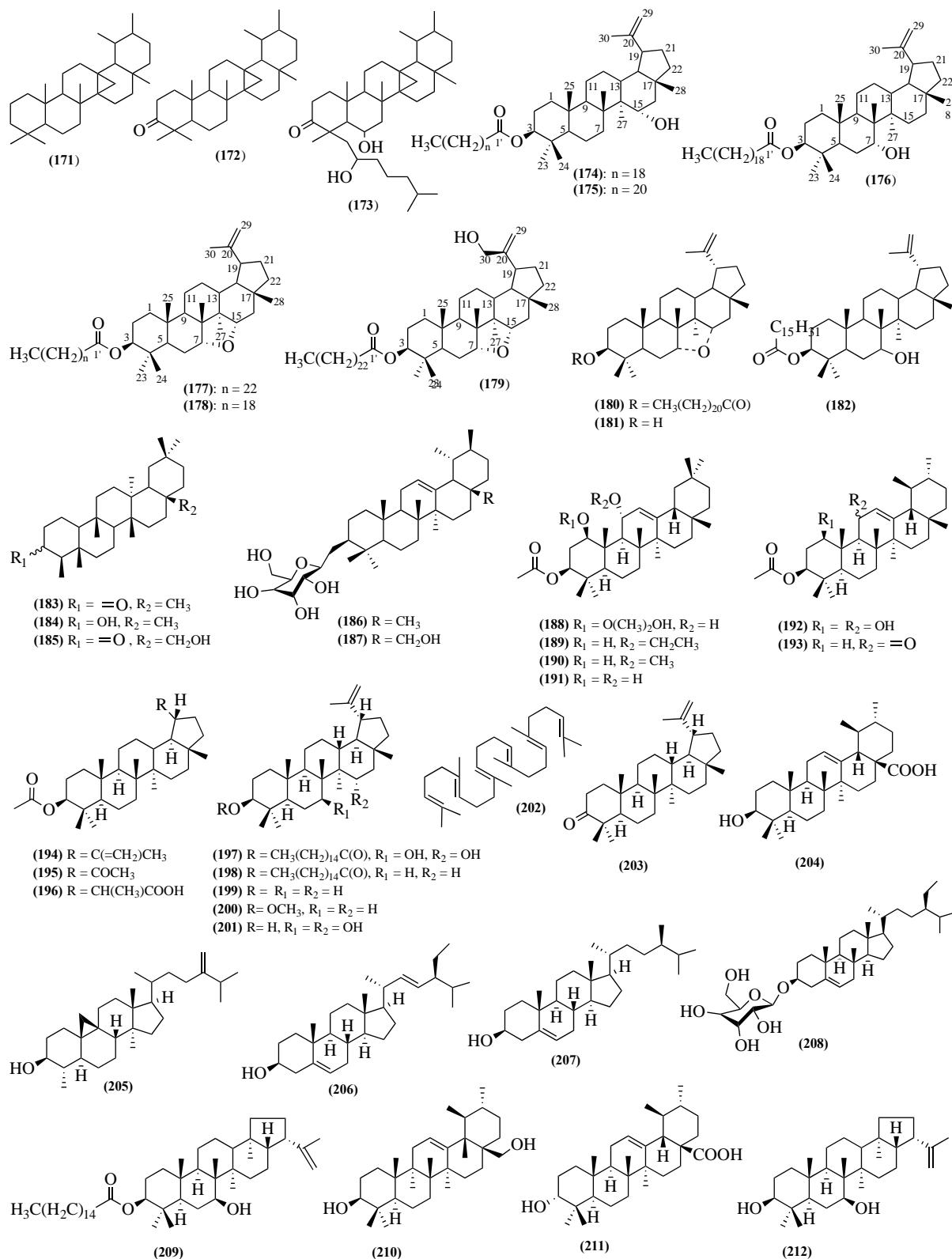


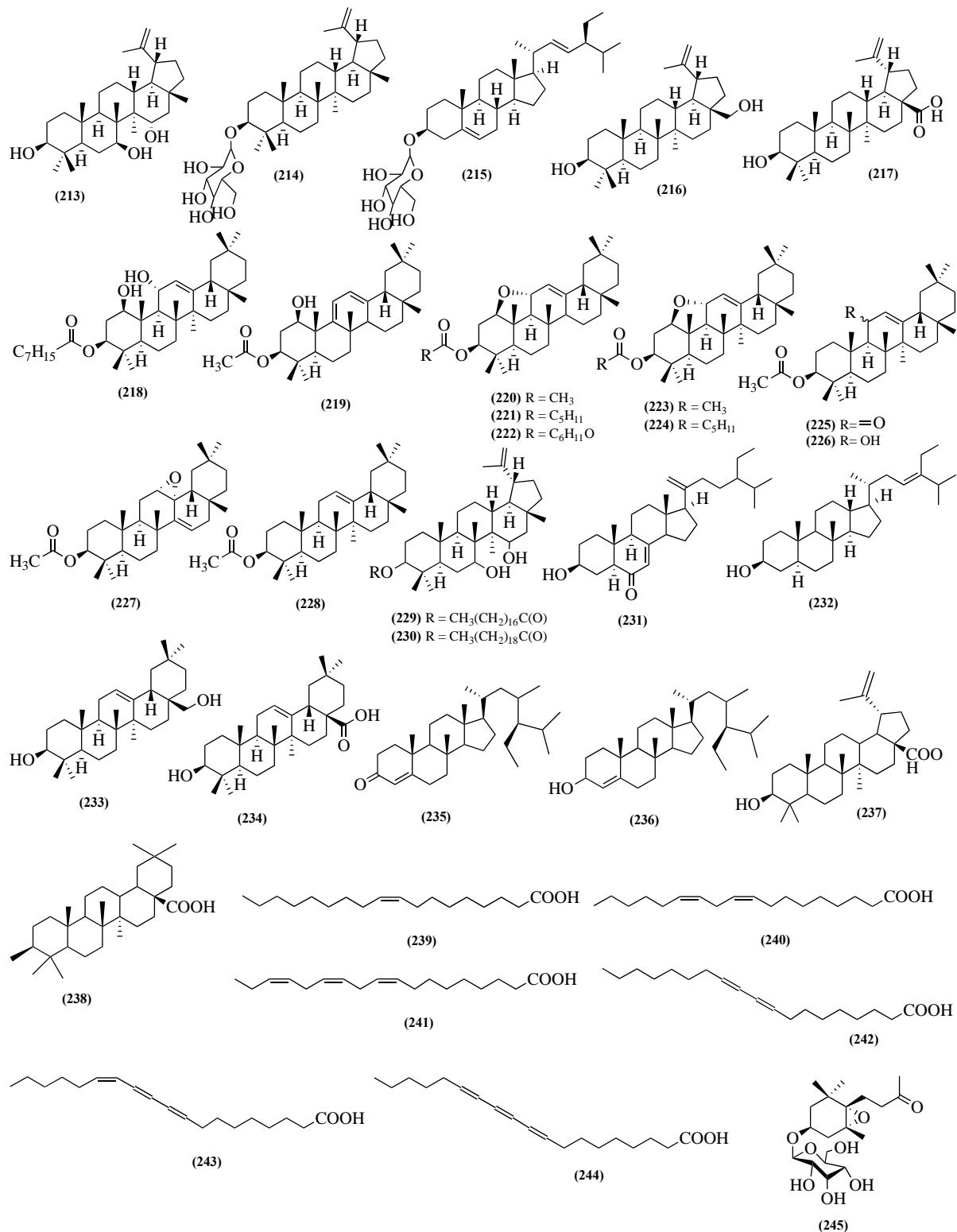


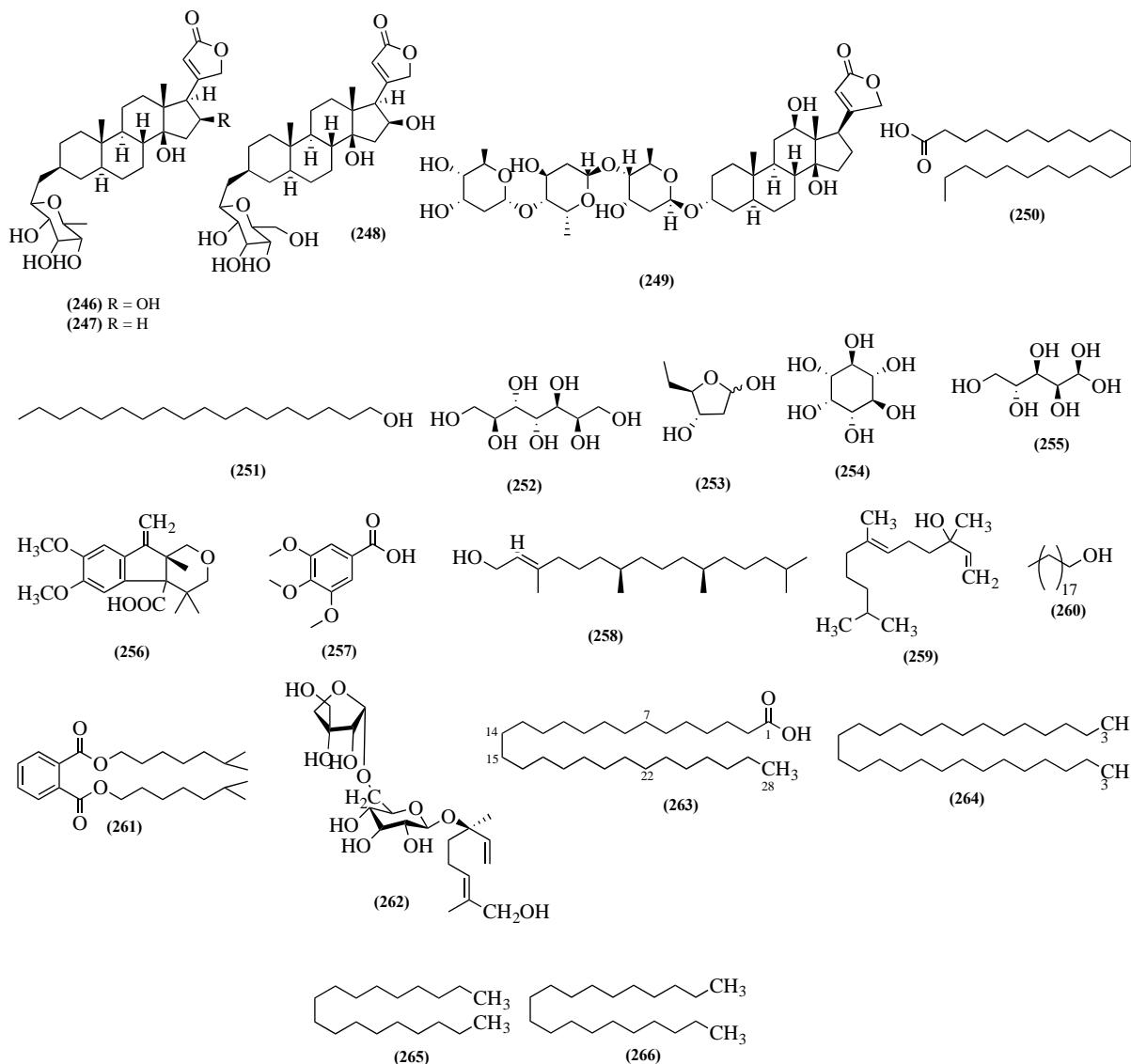












4. CONCLUSION

This compilation includes the phytochemicals, species and the host plants of the family Loranthaceae, and specifically provides some exploration of the literatures published. Some members of the family are reported to possess medicinal properties and are used to treat various ailments. Quercetin 3-*O*- α -L-rhamnopyranoside (Quercitrin) (**6**) is the major constituent of this family, and is important chemotaxonomic marker of the Loranthaceae plant species from a phytochemical point of view.

Declaration of Interest

I declare that there is no conflict of interest.

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REFERENCES

- Abdullahi MI, Musa AM, Muhammad KJ, Tajuddeen N, Sule MI. (2014). Isolation of two new hydroxyl-ethyl octalactones from the leaves of *Globimetula braunii* Mistletoe growing on *Piliostigma thonningii*. *Journal of Pharmaceutical Science and Pharmacy Practice*, 10, 200-203.
- Abubakar H, Musa AM, Abdullahi MI, Mzozoyana V, Yusuf AJ. (2020). Isolation and characterization of some flavonoids from the leaf of *Tapinanthus globiferus* growing on *Vitex doniana*. *Brazilian Journal of Biological Sciences*, 7, 239-245.
- Adesina SK, Illoh CH, Imoh IJ, Jacobs EI. (2013). African mistletoes (Loranthaceae); ethnopharmacology, chemistry and medicinal values: an update. *African Journal of Traditional and Complementary Alternative Medicine*, 10, 161–170.
- Al-Ghaithi F, Mandouh R, El-Ridi MR, Adeghate E, Amiri MH. (2004). Biochemical effect of *Citrullus colocynthis* in normal and diabetic rats. *Molecular and Cellular Biochemistry*, 261, 143-149.
- Anyam JV, Nvau JB, Thomas K, Eman S, Gray AI, Igoli JO. (2022). Dodoneine, its bicyclic lactone and a dihydroxyl-lupeol palmitate from *Tapinanthus globiferus*. *Tropical Journal of Natural Product Research*, 6, 938-942.
- Apaza TL, Serban AM, Cabanillas AH, Villacampa A, Rumbero A. (2019). Flavonoids of *Tripodanthus acutifolius* inhibit TNF- α production in LPS-activated THP-1 and B16-F10 cells. *Journal of Ethnopharmacology*, 242, 112036.
- Ayoola MD, Oriola AO, Faloye KO, Aladesanmi AJ. (2020). Two antihyperglycaemic compounds from *Globimetula braunii* (Engl.) Van Tiegh (Loranthaceae). *GSC. Biological and Pharmaceutical Science*, 11, 46-54.
- Bashar A, Juvik OJ, Dupont F, Francis GW, Fossen T. (2012). Novel aminoalkaloids from European mistletoe (*Viscum album* L.). *Phytochemistry Letters*, 5, 677-681.
- Beutler JA. (2009). Natural products as a foundation for drug discovery. *Current Protocol in Pharmacology*, 1, 9111-9121.
- Bick IRC. (1996). Alkaloids from Australian Flora. In: Pelletier, S.W. Editor. *Alkaloids: Chemical and Biological Perspectives*. Pergamon.
- Bo D, Yi D, Yun-Long H, Xiao-Meng W, Xue C, Xin-Sheng Y. (2013). Four new hemiterpenoid derivatives from *Taxillus chinensis*. *Fitoterapia*, 86, 1-5.
- Bruno NL, Joel TA, Jean RC, Makoa NA, Flora N, Gabriele P, Beate N, Hans GS, Catherin V, Silvere N, Norbert S. (2015). Two 2,6-dioxabicyclo[3.3.1]nonan-3-ones from *Phragmanthera capitata* (Spreng.) Balle (Loranthaceae). *Helvetica Chimica Acta*. 98, 945-952.
- Burkill HM. (1995). The useful plants of West Tropical Africa 2(3). Families J-L. Royal Botanic Gardens. Kew. 857 APG, 2003. An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG II. *Botanical Journal of the Linnean Society*, 141, 399-436.
- Burkill HM. (1985). The useful Plants of West Tropical Africa. 3 (families J-L) Royal Botanical Gardens, Kew: 548–560.
- Carmen MC, Miguel LL, Maria AA, Eduardo N, Juan T, Maria JAA. (2001). Cytotoxic diarylheptanoid from *Viscum cruciatum*. *Phytochemistry*, 58, 567-569.
- Cragg GM, Newman DJ. (2005). Biodiversity: a continuing source of novel drug leads. *Pure and Applied Chemistry*, 77, 7-24.
- Danladi S, Sule MI, Muhammad MA, Yaro AH. (2021). Central nervous system depressant activity of fractions of *Globimetula braunii* Engl. (Loranthaceae) growing on *Terminalia catappa* L.(Combretaceae) and isolation of lupeol. *Journal of Pharmaceutical and Biological Sciences*, 18, 196-206.
- Danladi S, Sule MI, Muhammad MA, Yaro AH. (2021). Isolation and characterization of some flavonoids from the leaves of *Globimetula braunii* (Loranthaceae) growing on *Terminalia catappa* L. (Combretaceae). *Tropical Journal of Natural Product Research*, 5, 2133-2136
- Deeni YY. (1989). Plants in Kano Ethnomedicine: Screening for Antimicrobial Activity and Alkaloids. M.Sc. thesis, Bayero University, Kano, Nigeria.
- Deliorman D, Calis I, Ergun F. (2001). A new acyclic monoterpane glucoside from *Viscum album* ssp. *album*. *Fitoterapia*. 72, 101-105.
- Edwin OO, Mohd PK, Patience OO, Deepshikha T, Mohammad FK, Kapil D, Rakesh M, Naibedya C. (2014). Analysis of constituents of the eastern Nigeria mistletoe, *Loranthus micranthus* linn revealed presence of new classes of osteogenic compounds. *Journal of Ethnopharmacology*, 151, 643-651.
- Edwin OO, Patience OO, Chukwuemeka SN, Justina NN, Wilfred OO, Akira K, Charles OE, Peter PA. novel sesquiterpene acid and an alkaloid from leaves of the Eastern Nigeria mistletoe, *Loranthus micranthus* with potent immunostimulatory activity on C57BL6 mice splenocytes and CD69 molecule. *Pharmaceutical Biology*, 49, 1271-1276.

- Elujoba AA. (1997). The role of pharmacognosy in phytotherapy, the challenges of our time. *Nigerian Journal of Natural Product*, 2, 34-36.
- Francoise LD, Sophie T, Didier F, Joel B. (2002). Flavonols from *Scurrula ferruginea* Danser (Loranthaceae). *Zeitschrift Naturforschung C Journal of Bioscience*, 7, 1092-1095.
- Fukunaga T, Nishiya K, Kajikawa I, Takeya K, Itokawa H. (1989). Studies on the constituents of Japanese mistletoes from different host trees, and their antimicrobial and hypotensive properties. *Chemical and Pharmaceutical Bulletin*, 37, 1543-1546.
- Gangwar JP, Saxena PN. (2010). Chemical constituents of *Dendrophoe falcate*. *University Bucur*, 19, 31-34.
- Gill LS, Onyibe HI. (1990). Mistletoes on rubber trees in Nigeria. Haustorium. *Journal of Sustainable Agriculture*, 1, 13-18.
- Goda MS, Elhady SS, Nafie MS, Bogari HA, Malatani RT, Hareeri RH, Badr JM, Donia MS. (2022). *Phragmanthera austroarabica* A.G. Mill. and J.A. Nyberg Triggers Apoptosis in MDA-MB-231 Cells in Vitro and in Vivo Assays: Simultaneous Determination of Selected Constituents. *Metabolites*, 12, 921.
- Goodman JWV. (2001). The Story of Taxol: Nature and Politics in the Pursuit of an Anti-Cancer Drug. *British Medical Journal*, 323, 115.
- Gurib-Fakim A. (2006). Medicinal plants: traditions of yesterday and drugs of tomorrow, *Molecular Aspects of Medicine*, 27, 1-93.
- Haizhen L, Zhun H, Chao L, Yao Z, Tao S, Qingwen H, Dongmei R. (2015). Three pairs of diastereoisomeric flavanone glycosides from *Viscum articulatum*. *Fitoterapia*, 102, 156-162.
- Harmine M, Pierre M, Sandra LF, Sylvain KD, Hayato I, Hiroshi N, Ephrem AN. (2017). Triterpenoids from seeds of *Tapinanthus bangwensis*. *Phytochemistry Letters*, 19, 23-29.
- Harvey AL. (2008). Natural products in drugs discovery. *Drug Discovery Today*, 13, 894-901.
- Hasan SM, Ahmed IM, Mondal S, Uddin SJ, Masud MM, Sadhu SK, Ishibashi M. (2006). Antioxidant. Antioxidant, antinociceptive activity and general toxicity study of *Dendrophoe falcata* and isolation of quercitrin as the major component. *Oriental Pharmacy and Experimental Medicine*, 6, 355-360.
- Hassan LG, Liman MG, Mshelia HE, Ogbiko C, Babagana A, Andrew O. (2018). Lupeol Acetate Isolated from n-Hexane Extract of *Tapinanthus globiferus* Leaf. *ChemSearch Journal*, 9, 83-88.
- Huaxing Q, Micheal GG. (2004). Loranthaceae. Flor. China. 5, 220.
- Hui Y, Zhi-Xin L, Qiong W, Guang-Qing L, Zhi-Jun L, Dao-Feng C, Jia-Kuan C, Tong-Shui Z. (2006). Antioxidative Flavanone Glycosides from the Branches and Leaves of *Viscum coloratum*. *Chemical and Pharmaceutical Bulletin*, 54, 133-135.
- Hussain HSN, Karatela YY. (1989). Traditional medicinal plants used by Hausa Tribe of Kano State of Nigeria. *International Journal of Crude Drug Research*, 27, 211-216.
- Ibrahim JA, Ayodele AE. (2013). Taxonomic Significance of Leaf Epidermal Characters of the Family Loranthaceae in Nigeria. *World Applied Science Journal*, 24, 1172-1179.
- Ikome HN, Ayimele GA, Bouobouo PL, Yadang SAF, Lah FCW, Tedonkeu AT. (2020). Antioxidant activity of flavonoids from the leaves of *Tapinanthus pentagonia* (Loranthaceae). *Journal of Phytopharmacology*, 9, 202-208.
- Irvine FR. (1961). Woody plants of Ghana. Oxford University Press. London.
- Ja'afar KM, Shajarahtunnur J, Norazah B. (2017). Antioxidant Activity of Leaf Extracts of *Globimetula braunii* (Engler) van Tiegh Parasitizing on *Piliostigma thonningii* and *Parkia biglobosa*. *Jurnal Teknologi*, 79, 43-47.
- Ja'afar KM, Shajarahtunnur J, Norazah B, Mohd BB, Satyajit DS, Keith JF, Mathias OS. (2017). Lactones and Flavonoids from the Leaves of *Globimetula braunii*. *Natural Product Communication*, 12, 1455-1458.
- Jeffreys D. (2005). Aspirin: The Remarkable Story of a Wonder drug: Bloomsbury Publishing PLC.
- Jia-Kun D, Duo C, Cui-Hu L, Jing G, Meng-Qing L, Na F, Ya-Hui W, Zheng-Liang S, Meng-Yang H. (2019). Three new bioactive flavonoid glycosides from *Viscum album*. *Chinese Journal of Natural Medicine*, 17, 545-550.
- Jun Y, Xiaokang W, Zhihui L, Shigetoshi K. (2007). A new tetrahydrofuran lignan glycoside from *Viscum coloratum*. *Asian Journal of Traditional Medicine*, 2, 86-88.
- Kazuyoshi O, Hendig W, Mutsuko M, Masahiro I, Made SP, Partomuan S, Hirotaka S. (2003). Indonesian Medicinal Plants. XXV.¹⁾ Cancer Cell Invasion Inhibitory Effects of Chemical Constituents in the Parasitic Plant_{SEP}^T*Scurrula atropurpurea* (Loranthaceae). *Chemical and Pharmaceutical Bulletin*, 51, 343-345.
- Kinghorn AD, Chin YW, Swanson SM. (2009). Discovery of natural product anticancer agents from biodiverse organisms. *Current Opinion in Drug Discovery and Development*, 12, 189-196.
- Kolb TE. (2002). Ecología del parasitismo en reino vegetal. In Plantasparasitas del la Peninsula Iberica e Islas Baleares Madrid, Edds. Saez-Lopez, J.A. P. Catalan and L. Saez. Mundi-Prensa.
- Kumar S, Prasad AK, Mehta SB. (2022). Isolation and structure elucidation of novel compounds from stem of *Dendrophoe falcate*. *Indian Journal of Chemistry*, 61, 999-1005.

- Lie-Chwen L, Tung-Hu T, Shao-Chieh S. (2011). A novel 8.O.6'-neolignan from *Taxillus theifer*. *Natural Product Research*, 25, 1319–1323.
- Lim YC, Rajabalaya R, Shirley HFL, Tennakoon KU, Quang-Vuong L, Idris A, Zulkipli, IN, Keasberry N, David SR. (2016). Parasitic mistletoes of the genera *Scurrula* and *Viscum*: from bench to bedside. *Molecules*, 21, 1048.
- Lin J, Lin Y. (1999). Flavonoids from the Leaves of *Loranthus kaoi* (Chao) Kiu. *Journal of Food and Drug Analysis*, 7, 185–190.
- Liyuan Y, Jun L, Bin Z, Yangang L, Baoquan Z. (2017). Activity of compounds from *Taxillus sutchunensis* as inhibitors of HCV NS3 series protease. *Natural Product Research*, 31, 487-491.
- Maher S, Fayyaz S, Naheed N, Dar Z. (2021). the isolation and screening of the bioactive compound of *Viscum album* against *Meloidogyne incognita*. *Pakistan Journal of Nematology*, 39, 46-51.
- Mallavadhani UV, Narasimhan K, Sudhakar AVS, Mahapatra A, Li W, van Breemen RB. (2006). Three new pentacyclic triterpenes and some flavonoids from the fruits of an Indian Ayurvedic plant *Dendrophthoe falcata* and their estrogen receptor binding activity. *Chemical and Pharmaceutical Bulletin*, 54, 740-744.
- Matthes H, Schad F, Buchward D, Schenk G. (2005). Endoscopic ultrasound-guided fine needle injection of *V. album* L. (mistletoe; *Helixor M*) in the therapy of primary inoperable pancreas cancer; a pilot study. *Gastroenterology*, 128, 988.
- Matthias OA, Chukwuemeka SN, Festus BCO, Patience OO. (2014). Isolation and structure elucidation of polyphenols from *Loranthus micranthus* Linn. parasitic on *Hevea brasiliensis* with anti-inflammatory property. *EXCLI Journal*, 13, 859-868.
- Maurice O, Helene C, Clarisse V, Jocelyn B, Guy R, Innocent-Pierre G, Christian C, Frederic B, Daniel P, Alain C, Jerome M, Jean-Marie C. (2007). Structure Elucidation of a Dihydopyranone from *Tapinanthus dodoneifolius*. *Journal of Natural Product*, 70, 2006-2009.
- Mfotie Njoya E, Hermine LD, Mkounga MP, Koert U, Nkengfack AE, McGaw LJ. (2020). Selective cytotoxic activity of isolated compounds from *Globimetula dinklagei* and *Phragmanthera capitata* (Loranthaceae). *Zeitschrift Naturforschung C Journal of Bioscience*, in press.
- Mkounga P, Maza DHL, Ouahouo WBM, Tyon NL, Hayato I, Hiroshi N, Nkengfack AE. (2016). New lupane-type triterpenoid derivatives from *Globimetula dinklagei* (Loranthaceae), a hemiparasitic plant growing on *Manihot esculenta* (Euphorbiaceae). *Zeitschrift Naturforschung C Journal of Bioscience*, 10, 0215-0235.
- Muhammad KJ, Jamil S, Basar N, Arriffin NM, Idris MT, Jibril S, Akanji FT. (2022). Antioxidant, Antimicrobial, and Antityrosinase Activities of Phytochemicals from the Leaves of *Globimetula braunii* (Engler) van Tiegh (Loranthaceae). *Bulletin of Chemical Society of Ethiopia*, 36, 387-397.
- Muhammad KJ, Jamil S, Basar N, Magaji MG. (2019). Anticonvulsant studies on the isolated compounds from the leaves of *Scurrula parasitica* L (Loranthaceae). *Malaysian Journal of Fundamental and Applied Science*, 15, 806-810.
- Muhammad KJ, Jamil S, Basar N, Sarker SD, Mohammed MG. Globrauneine A-F: (2020). Six new triterpenoid esters from the leaves of *Globimetula braunii*. *Natural Product Research*, 34, 2746-2753.
- Muhammad KJ, Jamil S, Basar N. (2019). Phytochemical study and biological activities of *Scurrula parasitica* L (Loranthaceae) leaves. *Journal of Research in Pharmacy*, 23, 522-531.
- Musa MA, Abdullahi IM, Kamal MJ, Magaji GM. (2014). Phytochemical screening and anticonvulsant studies of ethyl acetate fraction of *Globimetula braunii* on laboratory animals. *Asian Pacific Journal of Tropical Biomedicine*, 4, 285-289.
- Na H, Ting H, Yi-Chun W, Jun Y, Shigetoshi K. (2011). Flavanone Glycosides from *Viscum coloratum* and Their Inhibitory Effects on Osteoclast Formation. *Chemistry and Biodiversity*, 8, 1682-1688.
- Newman DJ, Cragg GM. (2012). Natural products as sources of new drugs over the 30 years from 1981 to 2010. *Journal of Natural Product*, 23, 311-335.
- Nguyen XN, Phan VK, Chau VM, Nanyoung K, SeonJu P, Hwa YL, Eun SK, Young HK, Sohyun K, Young-Sang K, Seung HK. (2013). Diarylheptanoids and flavonoids from *Viscum album* inhibit LPS-stimulated production of pro-inflammatory cytokines in bone marrow-derived dendritic cells. *Journal of Natural Product*, 76, 495-502.
- Noman OM, Nasr FA, Mothana RA, Alqahtani AS, Qamar W, Al-Mishari AA, Al-Rehaily AJ, Siddiqui NA, Alam P, Almarfadi OM. (2020). Isolation, Characterization, and HPTLC-Quantification of Compounds with Anticancer Potential from *Loranthus Acaciae* Zucc. *Separations*, 7, 43.
- Nonato de Oliveira Melo M, Oliveira AP, Wieckowski AE, Carvalho RS, Castro JC, Gomes de Oliveira FA, Gualberto Pereira HM, Feo da Veiga V, Capella MMA, Rocha L, Holandino C. (2018). Phenolic compounds from *Viscum album* tinctures enhanced antitumor activity in melanoma murine cancer cells. *Saudi Pharmaceutical Journal*, 26, 311-322.
- Obatomi DK, Aina VO, Temple VJ. (1996). Effects of African Mistletoe extract on Blood pressure in spontaneously hypersensitive rats. *International Journal of Pharmacognosy*, 34, 124-127.
- Obatomi DK. (1994). Anti-diabetic properties of the African mistletoes in treptozotozin-induced diabetic rats.

- Journal of Ethnopharmacology*, 43, 13-17.
- Ogbulie JN, Ogueke CC, Okorondou S. (2004). Antibacterial properties of *A. cordifolia*, *M. flurum*, *U. chaeme*, *B. pinnatum*, *C. albidem* and *A cilata* on some hospital isolates. *Nigerian Journal of Microbiology*, 18, 249-255.
- Ogechukwu OE, Ogoamaka OP, Sylvester NC, Hassan A, Debbab A, Okechukwu EC, Kawamura A, Peter P. (2011). Steroids and triterpenoids from Eastern Nigeria mistletoe, *Loranthus micranthus* Linn. (Loranthaceae) parasitic on *Kola acuminata* with immunomodulatory potentials. *Phytochemistry Letters*, 4, 357-362.
- Okigbo RN, Mbajiuka CO. (2005). Antimicrobial potentials of *Xylopia aethiopica* (Uda) and *Ocimum gratissimum* on some pathogens of man. *International Journal of Molecular Medicine and Advance Science*, 1, 392-397.
- Okpuzor J, Ogbunugafor HA, Kareem GK. (2009). Hepatic and hematologic effects of fractions of *Globimetula sebraunii* in normal albino rats. *International Journal of Experimental and Clinical Science*, 8, 182-189.
- Olagunju JA, Ismail FI, Gbile ZO. (1999). The hypoglycaemic property of Normal saline leaf of *Globimetula braunii* in alloxanised diabetic albino rats. *Biomedical Letters*, 60, 83-89.
- Oliver I. (1987). Teemoohlware- a refreshing bush tea. *Veld and Flora*, 73, 16.
- Omeje EO, Osadebe PO, Akira K, Amal H, Abdessamad D, Esimone CO, Nworu CS, Nwodo N, Proksch P. (2012). Three (-)-catechin-O-rhamnosides from the eastern Nigeria mistletoe with potent immunostimulatory and antioxidant activities. *Biomolecules*, 1, 1-6.
- Oriola AO, Aladesanmi AJ, Idowu TO, Akinwumi FO, Obuotor EM, Idowu T, Oyedeji AO. (2021). Ursane-type triterpenes, phenolics and phenolic derivatives from *Globimetula braunii* Leaf. *Molecules*, 26, 6528.
- Ouedraogo S, Ranaivo HR, Ndiaye M, Kabore ZI, Guissou PI, Bucher B. (2004). Cardiovascular properties of aqueous extract from *Mitragyna inermis* (wild). *Journal of Ethnopharmacology*, 93, 345-50.
- Patrick-Iwuanyanwu KC, Onyeike EN, Adhikari A. (2014). Isolation, identification and characterization of gallic acid derivatives from leaves of *Tapinanthus bangwensis*. *Journal of Natural Product*, 7, 14-19.
- Phillipson JD, Wright CW. (1991). Can ethnopharmacology contribute to the development of antimalarial agents? *Journal of Ethnopharmacology*, 32, 155-65.
- Puneetha GK, Amruthesh KN. (2016). Phytochemical screening and in vitro evaluation of antioxidant activity of various extracts of *Scurrula parasitica*. *International Journal of Pharmaceutical and Biological Science*, 6, 77-86.
- Quan-Yu L, Fei W, Lei Z, Jie-Ming X, Lia P, Yong-Hong ZA. (2015). Hydroxylated lupeol-based triterpenoid ester isolated from the *Scurrula parasitica* Parasitic on *Nerium indicum*. *Helvetica Chimica Acta*, 98, 627-632.
- Quan-yu L, Fei W, Yong-Hong Z, Feng NI. (2016). Chemical constituents of *Scurrula paracitica*. *China Journal of Chinese Materia Medica*, 41, 3956-3561.
- Robert B, Monica J, Bernard TK, Oyvind MA. (2011). Primitive anthocyanin from flowers of three hemiparasitic African mistletoes. *School Research Library*, 3, 1-5.
- Rui JL, Guan EY, Hong JB, Qiong Z, Jian KL, Qing SL, Zhao MZ. (2012). A new flavonoids glycosides from mistletoe transformed by rhodobacter sphaeroides. *Chemistry of Natural Compound*, 48, 682-684.
- Shen C, Chang Y, Ho L. (1993). Nuclear magnetic resonance studies of 5,7-dihydroxy Flavonoids. *Phytochemistry*, 34, 843-845.
- Sher H, Alyemeni MN. (2011). Pharmaceutical important plants used in traditional system of Arab medicine for the treatment of livestock ailments in the kingdom of Saudi Arabia. *African Journal of Biotechnology*, 10, 9153-9159.
- Shoeb M. (2006). Anticancer agents from medicinal plants. *Bangladesh Journal of Pharmacology*, 1, 35-41.
- Soberón JR, Sgariglia MA, Sampietro DA, Quiroga EN, González SM, Vattuone MA. (2010). Free radical scavenging activities and inhibition of inflammatory enzymes of phenolics isolated from *Tripodanthus acutifolius*. *Journal of Pharmacology*, 130, 329-333.
- Takashi I, Etsuji T, Hendig W, Kazuyoshi O, Hirotaka S. (2001). A complex of perseitol and K⁺ ion from *Scurrula fusca* (Loranthaceae). *Tetrahedron Letters*, 42, 6887-6889.
- Tarfa FD, Igoli JO, Gray AI, Adoga GI, Gamaniel KS. (2022). Characterization of potential hypoglycaemic agents from *Tapinanthus sessilifolius* Parasitic on *Psidium guajava*. *Journal of Phytomedicine Therapy*, 21, 1003
- Ticona LA, Estrada CT, Sánchez AR. (2020). Inhibition of melanin production and tyrosinase activity by flavonoids isolated from *Loranthus acutifolius*, *Natural Product Research*, in press.
- Traore F, Gasquet M, Laget M, Guiraud H, Giorgio CD, Azas N, Doumbo O, Timon David P. (2000). Toxicity and genotoxicity of antimalarial alkaloid rich extracts derived from *Mitragyna inermis*, *O. kuntze* and *Nuclea latifolia*. *Phytotherapy Research*, 4, 608-11.
- Tung-Hu T, Yi-Chen L, Lie-Chwen L. (2010). Flavonoids from *Taxillus theifer*. *Journal of Food and Drug Analysis*, 18, 256-262.
- UNESCO. (1998). FIT/504-RAF-48. Terminal Report: Promotion of Ethnobotany and the sustainable use of plant

- resources in Africa. United Nations Educational, Science and Cultural organization (UNESCO).
- Wong DZH, Kadir HA, Ling SK. (2012). Bioassay-guided isolation of neuroprotective compounds from *Loranthus parasiticus* against H₂O₂-induced oxidative damage in NG108-15 cells. *Journal of Ethnopharmacology*, 139, 256-264.
- World Health Organization. (2014). Traditional Medicine. Geneva.
- Yang L, Yan-Li Z, Yong-Ping Y, Xiao-Li L. (2011). Chemical constituents of *Viscum album* var. *meridianum*. *Biochemical Systematic and Ecology*, 39, 849-852.
- Young-Kyo K, Young SK, Sang UC, Shi YR. (2004). Isolation of flavonol rhamnosides from *Loranthus tanakae* and cytotoxic effect of them on human tumor cell lines. *Archives of Pharmacal Research*, 27, 44-47.